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Synthesis and Biological Evaluation of Novel cYY Analogues targeting *Mycobacterium tuberculosis* CYP121A1

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Abstract

The rise in multidrug resistant (MDR) cases of tuberculosis (TB) has led to the need for the development of TB drugs with different mechanisms of action. The genome sequence of *Mycobacterium tuberculosis* (*Mtb*) revealed twenty different genes coding for cytochrome P450s. CYP121A1 catalyzes a C-C crosslinking reaction of dicycloytyrosine (cYY) producing mycocyclosin and current research suggests that either mycocyclosin is essential or the overproduction of cYY is toxic to *Mtb*. A series of 1,4-dibenzyl-2-imidazol-1-yl-methylpiperazine derivatives were designed and synthesised as cYY mimics. The derivatives substituted in the 4-position of the phenyl rings with halides or alkyl group showed promising antimycobacterial activity (MIC 6.25 µg/mL), with the more lipophilic branched alkyl derivatives displaying optimal binding affinity with CYP121A1 (ⁱPr K_D = 1.6 µM; ^tBu K_D = 1.2 µM). Computational studies revealed two possible binding modes within the CYP121A1 active site both of which would effectively block cYY from binding.

Keywords

CYP121A1; *Mycobacterium tuberculosis*; 1,4-dibenzyl-2-imidazol-1-yl-methylpiperazine derivatives; binding affinity assays; molecular modelling

1. Introduction

Tuberculosis (TB) is the ninth leading cause of death worldwide, ranking above Human Immunodeficiency Virus (HIV/AIDS). In 2017, the estimated TB deaths were 1.3 million among HIV-negative people and 300,000 among HIV-positive people, and the estimated incident TB cases were 10.0 million worldwide.¹ The standard therapeutic regimens involve the combination of four first-line drugs, often isoniazid, rifampicin, pyrazinamide and either streptomycin or ethambutol, depending upon whether there is a latent or active type of infection.² Although the current medications for treating drug-sensitive TB are effective when there is optimum adherence of patients, a common problem in real-life conditions appears with TB-infected patients who generally do not adhere strictly to dosages.^{3,4}

The duration for treating drug-sensitive TB is usually 6 months with the four most effective agents from the first-line oral drugs administered in a single prescription for the first two months of treatment, and two of the four taken for a subsequent four months in the continuation phase, resulting in patient adherence issues.⁵ When administered in sub-optimum conditions, chronic cases of infectious drug-resistant TB appear. Long treatment durations are usually required because *Mycobacterium tuberculosis* (*Mtb*), the causative pathogen, can develop a dormancy phenotype under nutrient depletion and anaerobiosis conditions which is tolerant to several anti-TB drugs.^{6,7} The development of novel anti-TB drugs has become a priority in view of the increasing global incidence of strains resistant to at least rifampicin and isoniazid (MDR-TB), or to rifampicin, isoniazid and one of the injectable second-line anti-TB drugs as well as to any of the fluoroquinolone drug series (XDR-TB),¹ in addition to the interactions between anti-TB drugs and antiretroviral medications, and the need to treat latent TB-infected patients before the bacteria transform into their active form.⁸ Studies following the unraveling of the genome sequence of the virulent *Mtb* H37Rv

strain revealed numerous genes with unknown functions.⁹ Among these were twenty different cytochrome P450 (CYP) genes coding for P450s.¹⁰ This large number of CYP genes was uncommon for a bacterium and, as a result, studies over the past twenty years have focused on characterizing these *Mtb* P450s, several of which play crucial roles in the survival of *Mtb*. Subsequent genome studies with *M. smegmatis* revealed several P450s, confirming the multiple important functions for the P450s in the actinobacteria.¹¹

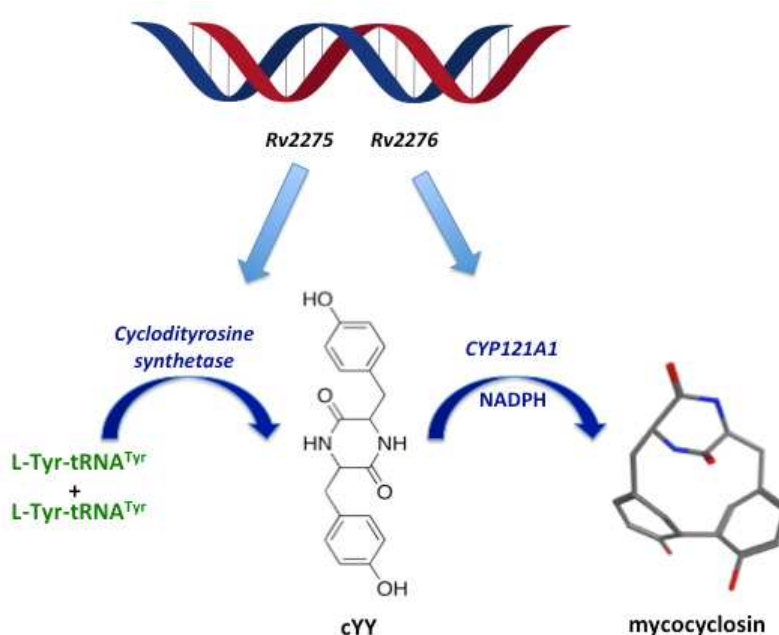


Figure 1. Cyclo-dityrosine synthetase and CYP121A1 reactions to produce the cyclic dipeptide, cyclo-*L*-Tyr-*L*-Tyr (cYY), and the novel secondary metabolite mycocyclosin.

CYP121A1 (mycocyclosin synthase) was shown to be essential for bacterial growth by *in vitro* gene knockout studies.¹²⁻¹⁴ Also, CYP121A1, along with CYP128A1 and CYP141A1, are the only *Mtb* P450s that appear to be conserved within members of the *Mtb* complex because their homologues do not appear in the genomes of the other members of the actinobacteria family. The first evidence of CYP121A1 function in *Mtb* was derived from its gene position, which is located in an operon harboring two

enzymes involved in the formation of cyclo-di-L-tyrosine (cYY). The first enzyme is a cyclodipeptide synthase (encoded by *Rv2275*), which uses aminoacyl-tRNA synthetases (L-tyrosyl-tRNA^{Tyr}) to catalyse the ATP-independent formation of cYY (Figure 1).^{15,16} Then, CYP121A1 (encoded by *Rv2276*) catalyzes a C-C crosslinking reaction between the respective carbons in the *ortho* position of the phenolic hydroxyl of cYY, producing mycocyclosin (Figure 1). *Rv2276* was found to be an essential *Mtb* gene, and it was suggested that either mycocyclosin was essential or the overproduction of cYY is toxic.^{14,17} Assays conducted on CYP121A1 utilising a ferredoxin and ferredoxin reductase system demonstrated that CYP121A1 catalyzes multiple turnovers of cYY in a complicated multistep process to form mycocyclosin as the single major product in the presence of NADPH.¹⁷

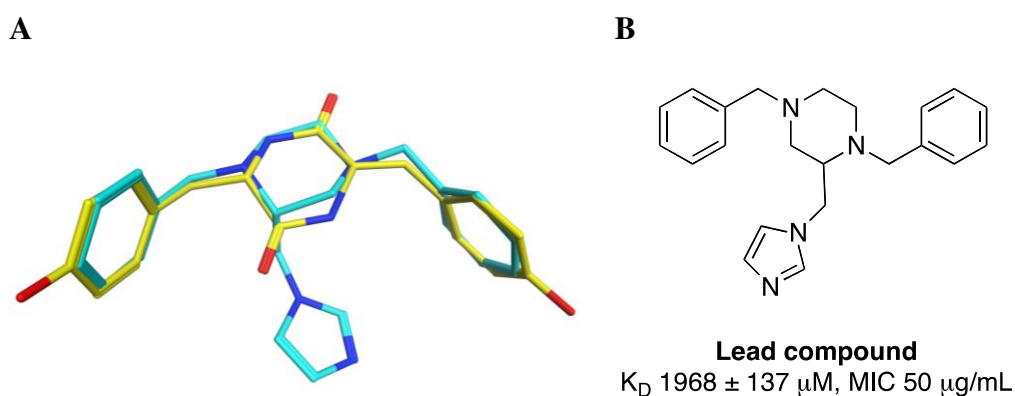


Figure 2. (A) Flexible alignment of cYY (yellow) and lead compound (cyan) showing overlap (B) Binding affinity and MIC data for lead compound.

In the development of CYP121A1 inhibitors, we have described several series of compounds with chemical scaffolds that mimic the natural substrate cYY.^{18,19} From these series 1,4-dibenzyl-2-imidazol-1-yl-methylpiperazine derivatives were identified as potential leads for further development.¹⁸ Flexible alignment of these derivatives with cYY showed good overlap (Figure 2) suggesting they would occupy a similar area as cYY within the CYP121A1 active site, while the addition of an azole haem-binding

The imidazole products (**8**) were obtained from a six-step synthetic route (Scheme 1). Substituted benzaldehyde derivatives (**1**) were reacted with ethylenediamine (**2**) in ethanol for 6 h at room temperature to give the di-Schiff base (di-imine) (**3**), which precipitated from the reaction as pure crystalline solids in high yields of 81-99%. The imine bond was reduced by NaBH₄ followed by careful aqueous work-up while cooling in an ice bath to give the diamines (**4**) in good yields (75-97%) with the exception of the pyridinyl derivatives (**4i** and **4j**), which were obtained in yields of 29 and 31% respectively owing to their water solubility. The synthesised diamines (**4**) on reaction with ethyl 2,3-dibromopropionate²⁰ resulted in the formation of ethyl 1,4-bis(substituted) piperazine-2-carboxylate (**5**) with yields in the range of 65-85%. Piperazine, as a six-membered cyclic system, can exist in both chair and boat conformations. The lowest energy and most stable conformation for the synthesised esters, using the conformational import algorithm²¹ in MOE²² followed by optimisation using the PM3 model Hamiltonian²³ is shown in Figure 3.

From the proposed 3D structure, the axial conformation for the ester moiety at C-2 was preferred,²⁴ this conformation of the ester intermediates was analysed by ¹H NMR and 2D COSY experiments. The H-2 proton had vicinal three-bond couplings across single bonds with both axial and equatorial H-3 protons, and appeared as doublet of doublets (dd) in the ¹H NMR spectra in derivatives **5a**, **5c**, **5e**, **5h** and **5i** with coupling constants in the range of 3.4-3.6 Hz for the vicinal equatorial-equatorial couplings, and larger values in the range of 5.5-6.0 Hz for the vicinal equatorial-axial couplings. In some derivatives, the H-2 proton couplings appeared as a multiplet (m) as seen in **5b** and **5d**, or interfered with other protons as seen in **5f**, **5g** and **5j**. Geminal spin-spin couplings were conserved between the H-7 protons and the H-8 protons, and

in the ^1H NMR spectra, each proton appeared as a doublet (d) with a geminal coupling constant in the range of 13.0-14.0 Hz.

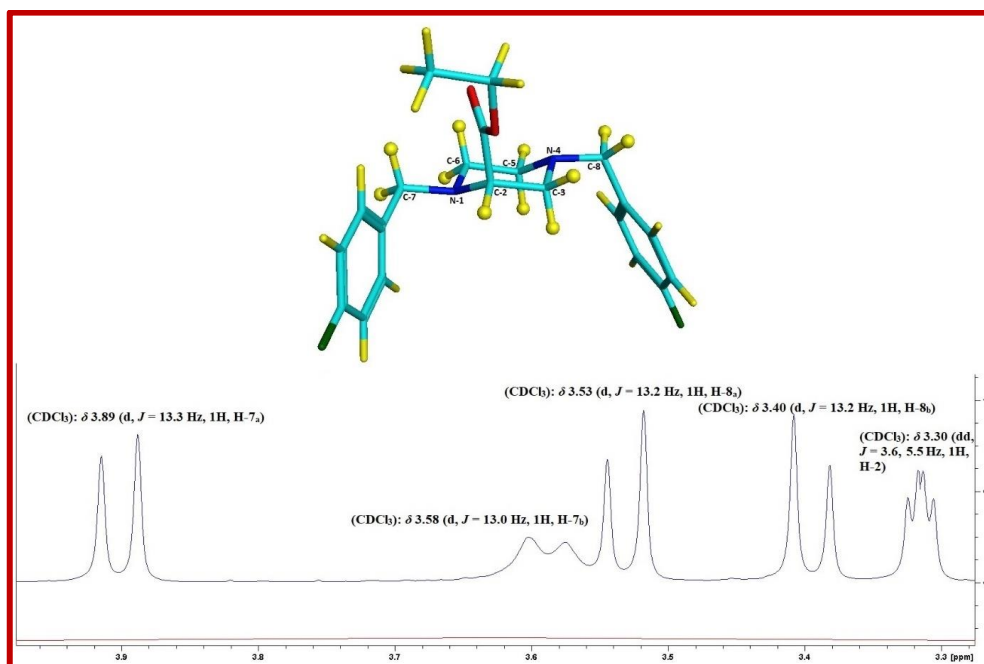
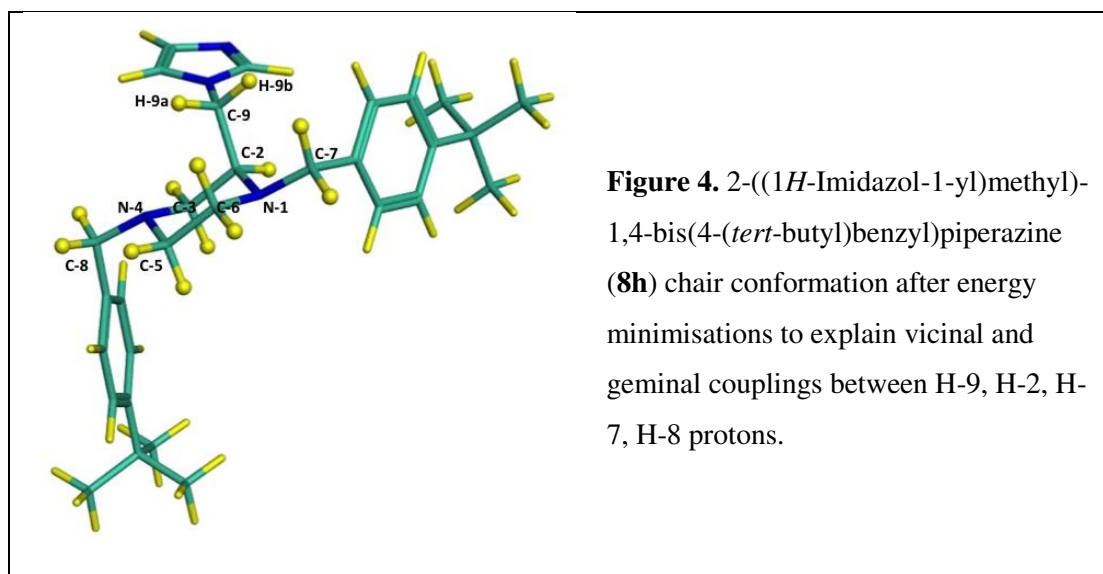


Figure 3. Ethyl 1,4-bis(4-fluorobenzyl)piperazine-2-carboxylate (**5a**) chair conformation with vicinal and geminal couplings of H-2, H-7 and H-8 protons.

The synthesised esters (**5**) were reduced to the corresponding primary alcohols (**6**) by LiAlH_4 after overnight stirring at room temperature with yields in the range of 56-91% obtained. Conversion of the alcohols (**6**) to the alkyl chlorides proceeded through a nucleophilic substitution $\text{S}_{\text{N}}2$ mechanism using thionyl chloride (10 equivalents) at room temperature for 48 h. A trial to accelerate the chlorination reaction rate was done by refluxing the synthesised alcohol with thionyl chloride (2 equivalents) at 80°C , but complex mixtures were obtained, and the products could not be separated or identified upon refluxing, especially for the halide derivatives (**7a-7d**). The yields of the chloride products (**7**) from reaction at room temperature for 48 h were in the range of 50-88%. The imidazolate ion, which acts as a nucleophile, was formed *in situ* by reacting imidazole with a base, for 1 h at 45°C in acetonitrile, followed by a $\text{S}_{\text{N}}2$ reaction

between the imidazolate and the synthesised alkyl chloride (7). When sodium hydride (60% dispersion in mineral oil) was used as a base according to the reported method,¹⁸ a complex mixture was obtained, and the product could only be separated in a very low yield (< 10%). Potassium carbonate was selected as the optimum base, and the reaction was complete after refluxing at 70 °C for 48 h with yields in the range of 51-69%.



A characteristic feature for the final imidazoles was the geminal and vicinal proton-proton coupling. The doublet of doublets (dd) at 4.28-4.29 ppm was assigned to the geminal ($J = 13.8\text{-}14.0$ Hz) and vicinal ($J = 4.3\text{-}4.4$ Hz) couplings of the H-9_a proton with H-9_b and H-2 protons, respectively. The doublet of doublets at 4.20-4.21 ppm was assigned to the geminal ($J = 13.8\text{-}14.0$ Hz) and vicinal ($J = 7.6\text{-}8.0$ Hz) coupling of the H-9_b proton with H-9_a and H-2 protons, respectively. The vicinal spin-spin coupling constants illustrated the difference in the chemical shifts between H-9_a and H-9_b protons. Vicinal spin-spin coupling constants were generally in the range of 6-8 Hz and, as the proton was closer to the imidazole group (electron-deficient group), smaller vicinal J values were noticed. From the proposed 3D-structure after energy minimisations (Figure 4), the H-9_a proton appeared to be closer to the imidazole group than the H-9_b proton. Consequently, the H-9_a proton was less shielded and its vicinal

coupling constant was less than 6.0 Hz while the H-9_b proton was more shielded and its vicinal coupling constant was within the range of 6-8 Hz.

2.2 CYP121A1 Ligand Binding Affinity

The CYP121A1 binding affinity (K_D) of the various compounds was determined by UV-vis optical titration.²⁵ Binding titrations were carried out for the synthesised imidazoles (**8**), which showed a type II (inhibitor-like) red shift in the haem Soret peak to a longer wavelength (Figure 5, for example **8h**) indicating that, in the solution state, most of these compounds coordinate either (i) directly to the CYP121A1 haem iron or (ii) indirectly to the haem iron through an interstitial water molecule.

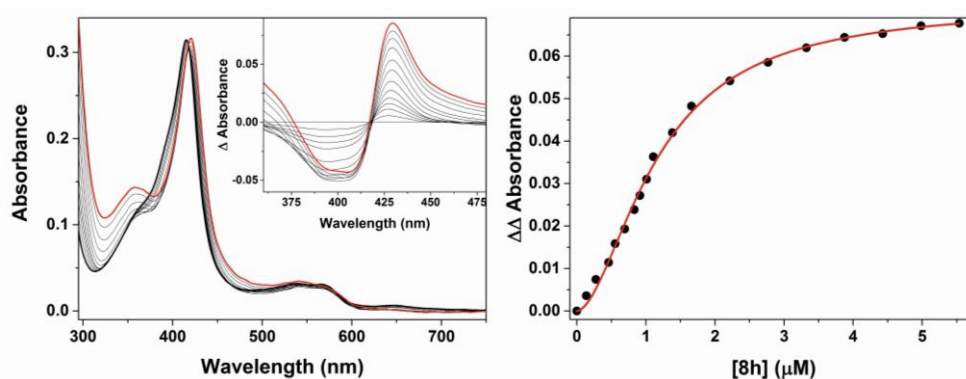
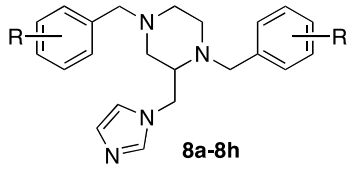


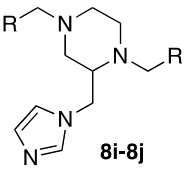
Figure 5. UV-Vis optical binding titration for compound **8h** binding to CYP121A1. The left hand panel shows data from a compound **8h** titration with CYP121A1 ($\sim 4.7 \mu\text{M}$) with the ligand-free spectrum as a thick black line, spectra following progressive additions of **8h** as thin solid lines, and the final near-saturated protein spectrum shown as a thick red line. The inset shows overlaid difference spectra generated by the subtraction of the starting spectrum from each consecutive ligand-bound spectrum collected in the titration. The right hand panel shows a plot of compound **8h**-induced absorbance change – calculated as the difference between the peak and trough in the difference spectra in the left hand panel, using the same wavelength pair (429 and 392 nm, respectively) throughout. Data were fitted using the Hill equation to give a **8h** K_D value of $1.2 \pm 0.1 \mu\text{M}$.

Halogenated derivatives (**8b-8d**) displayed moderate binding affinity with the best K_D value observed for the 3,5-diCl compound **8d**. The 4-F compound **8a** was noted to undergo photo- and thermal-decomposition in this assay so the data obtained was not reliable.

Table 1. Binding affinity (K_D) values for compounds against *M. tuberculosis* H37Rv



8a-8h



8i-8j

Compound	R	K_D (μM) ^a	Soret peak shift (nm)
Lead	H	1968 ± 137	416.5 to 417.5
8b	4-Cl	26.8 ± 7.9	416.5 to 419
8c	4-Br	11.9 ± 2.1	416.5 to 421
8d	3,5-diCl	8.1 ± 0.4	416.5 to 419
8e	4-OCH ₃	42.0 ± 14.3	416.5 to 419.5
8f	4-CH ₂ CH ₃	3.7 ± 0.1	416.5 to 423.5
8g	4-CH(CH ₃) ₂	1.6 ± 0.1	416.5 to 422.5
8h	4-C(CH ₃) ₃	1.2 ± 0.1	416.5 to 419.5
8i	Pyridin-4-yl	39.2 ± 7.9	416.5 to 419.5
8j	Pyridin-3-yl	489 ± 27	416.5 to 421.5
Fluconazole		8.6 ± 0.2	
Clotrimazole		0.07 ± 0.01	
cYY		5.8 ± 0.2	

^acompound **8a** (R = 4-F) noted to undergo photo- and thermal-decomposition during this assay.

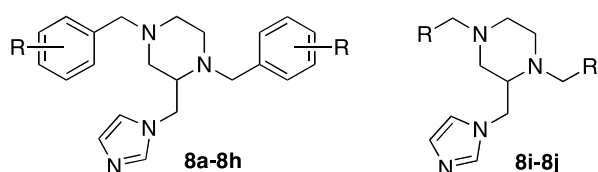
The lipophilic derivatives had the best binding of the imidazole nitrogen to the haem iron with K_D values of 3.7 ± 0.1 , 1.6 ± 0.1 and 1.2 ± 0.1 μM for **8f**, **8g** and **8h**, respectively, while the natural substrate cYY had a K_D value of 5.8 ± 0.2 μM . The

moderately hydrophilic 4-OMe (**8e**) and pyridines (**8i** and **8j**) had weak binding affinity to CYP121A1 (Table 1). Compounds **8f** and **8g** induce the most extensive Soret absorbance shifts (416.5 to 422.5/423.5 nm), suggesting that the CYP121A1 haem iron is predominantly coordinated by a direct imidazole nitrogen bond in these cases. Compounds **8b-8e** and **8h-8j** show less extensive Soret red shifts on binding the CYP121A1 haem, suggesting that ligation modes in these cases may involve imidazole nitrogen interactions with haem iron mediated through an interstitial water ligand.

2.3 MIC determination against *Mycobacterium tuberculosis*

The derivatives were screened against *M. tuberculosis* H37Rv by the REMA (Resazurin Microtiter Assay) method.²⁶ With the exception of the pyridyl compounds **8i** and **8j**, which were inactive (MIC >100 µg/mL), all the compounds were more active than the lead parent compound (MIC 50 µg/mL). In the halogenated series, the 4-monosubstituted benzene ring derivatives **8a-8c** showed good inhibitory activity (MIC 6.25 µg/mL) compared with the 3,5-dichloro substituted derivative **8d** (MIC 25 µg/mL). The alkyl substituted series all showed promising inhibitory activity (MIC 6.25 µg/mL), while the more hydrophilic 4-methoxy derivative (**8e**) was less effective (MIC 25 µg/mL).

Table 2. MIC values for compounds against *M. tuberculosis* H37Rv



Compound	R	Mol.Wt.	MIC ($\mu\text{g/mL}$)	cLogP ^a
Lead	H	346.469	50	3.56
8a	4-F	382.450	6.25	3.87
8b	4-Cl	415.359	6.25	4.67
8c	4-Br	504.261	6.25	5.21
8d	3,5-diCl	484.249	25	5.79
8e	4-OCH ₃	406.521	25	3.3
8f	4-CH ₂ CH ₃	402.575	6.25	5.36
8g	4-CH(CH ₃) ₂	430.628	6.25	6.02
8h	4-C(CH ₃) ₃	458.681	6.25	6.97
8i	Pyridin-4-yl	348.445	>100	0.88
8j	Pyridin-3-yl	348.445	>100	0.88
Fluconazole		306.271	>100	0.87
Clotrimazole		344.837	20	5.97
cYY		326.347	-	1.26

^a cLogP was determined using Crippen's fragmentation

2.4 Molecular Modelling

Molecular modelling of the compounds was performed using Molecular Operating Environment (MOE) software and the crystal structure of cYY co-crystallised with CYP121A1 (PDB 3G5H).¹⁷ Most of the compounds displayed two binding modes

when docked within the CYP121A1 active type, both of which show indirect binding with the haem via an interstitial water molecule.

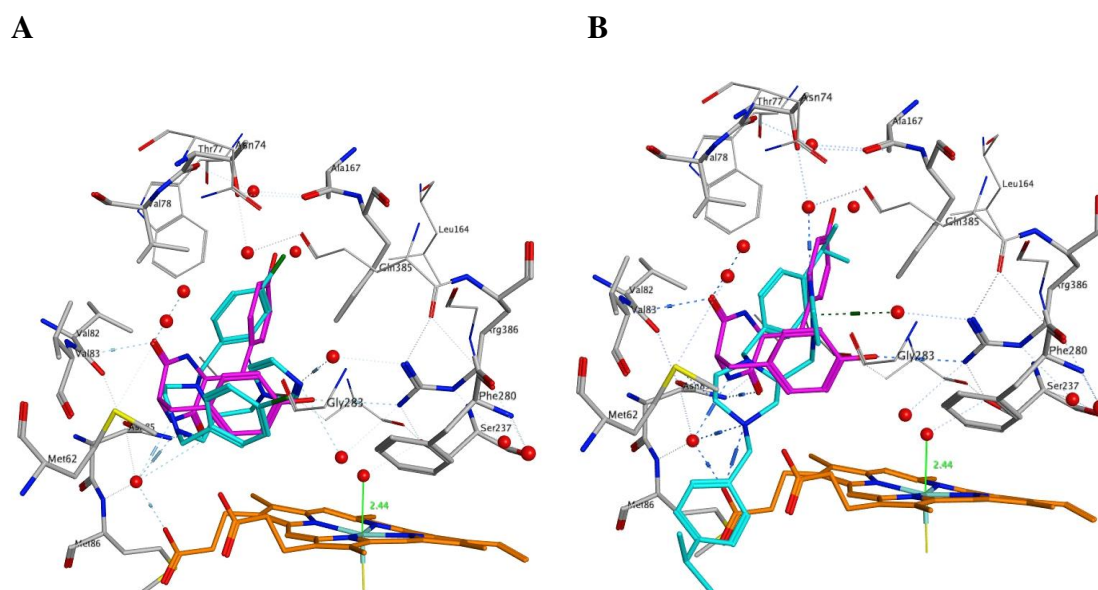


Figure 6. Docking of representative piperazines (cyan) (A) **8b** in binding mode 1 and (B) **8g** in binding mode 2 in the CYP121A1 active site compared with cYY (magenta). [Haem (orange), water molecules (red spheres)]

The first docking mode (mode 1) mimicked the cYY conformation as illustrated by the chloro derivative **8b** (Figure 6A). The most consistent binding interactions were observed between the imidazole ring and Arg386 via a water molecule and the protonated piperazine NH⁺ and Met86 effectively bridging the active site above the haem (Table 2). In the second binding mode (mode 2) a more open/flexible conformation was observed as illustrated by the isopropyl derivative **8g** (Figure 6B). This increased flexibility is possible owing to the piperazine ring which can exist in both chair and boat conformations. In mode 2, binding interactions were observed between the imidazole and a number of different amino acids (e.g. Asn74, Gly283, Gln385 and Arg386) via a water molecule with additional binding between the protonated piperazine NH⁺ and Met86. The pyridine derivatives **8i** and **8j** formed additional H-bonding interactions through the two pyridine rings. Multiple

hydrophobic interactions were observed between the piperazines (**8**) for both binding modes 1 and 2 including residues Met62, Thr77, Val78, Val82, Val83, Asn85, Leu164, Ala167, Phe168, Thr229, Ala233, Gly232, Phe280 and Leu284. For the unsubstituted lead compound and the fluoro (**8a**), chloro (**8b**) and pyridine (**8i** and **8j**) derivatives, binding mode 1 was energetically preferred (Table 2).

Table 2. Computational binding energies of piperazines **8** and CYP121A1

Compound	R	Mode 1 S-value (kcal/mol)	Mode 2 S-value (kcal/mol)
Lead	H	-4.69	-2.97
8a	4-F	-6.29	-4.61
8b	4-Cl	-4.53	-4.23
8c	4-Br	-	-2.78
8d	3,5-diCl	-	-3.64
8e	4-OCH ₃	-4.73	-4.79
8f	4-CH ₂ CH ₃	-3.34	-4.29
8g	4-CH(CH ₃) ₂	-3.18	-4.21
8h	4-C(CH ₃) ₃	-	0.04
8i	Pyridin-4-yl	-5.00	-2.35
8j	Pyridin-3-yl	-5.32/-6.58	-4.26

S-value = lowest binding energy

The methoxy derivative (**8e**) showed comparable S-values for binding modes 1 and 2, while the ethyl (**8f**) and isopropyl (**8g**) derivatives had improved S-values for binding mode 2 (Table 2). For the more bulky bromo (**8c**), dichloro (**8d**) and *tert*-butyl (**8h**) derivatives only binding mode 2 was observed with the *tert*-butyl derivative the least energetically favourable. Two possible mode 1 orientations were observed for the 3-pyridyl derivative (**8j**) with indirect interaction with the haem via a water molecule

either through imidazole (S-value -5.32) or through one of the pyridine rings (S-value -6.58.).

To investigate the binding modes further molecular dynamics simulations were run for 50ns for the CYP121A1 and isopropyl derivative (**8g**) and *tert*-butyl derivative (**8h**) complexes using the Desmond programme of Maestro.^{27,28} For the isopropyl (**8g**)-CYP121A1 complex, the RMSD changed from 1.2 Å at zero time to 2.1 Å at ~ 10 ns; then the protein was equilibrated with no evident RMSD fluctuations observed and the ligand aligned. For the *tert*-butyl (**8h**)-CYP121A1 complex, equilibrium and ligand alignment required a longer run time with the RMSD changing from 0.8 Å at zero time to 2.0 Å at ~ 30 ns. (Figure 7A). Protein interactions with the ligand were checked throughout the simulation and interactions ordered by their type; specifically hydrogen bonds, hydrophobic, ionic, and water bridges (Figure 7B). The isopropyl derivative (**8g**) formed more interactions over the course of the 50ns simulation compared with the *tert*-butyl derivative (**8h**). As illustrated in Figure 7C, water bridge interactions between the imidazole and Asp282 and Gln385, and the protonated piperazine and Val83 were significant for **8g** with additional hydrophobic interactions between the 4-*i*Pr-phenyl rings and Phe168 and Pro285. For **8h** the major contributor was a water bridge interaction between the imidazole ring and Arg386, with additional binding interactions between one of the *t*Bu-phenyl rings and His343 (π - π stacking) and Phe168 (hydrophobic) (Figure 7C).

iPr (8g)	tBu (8h)
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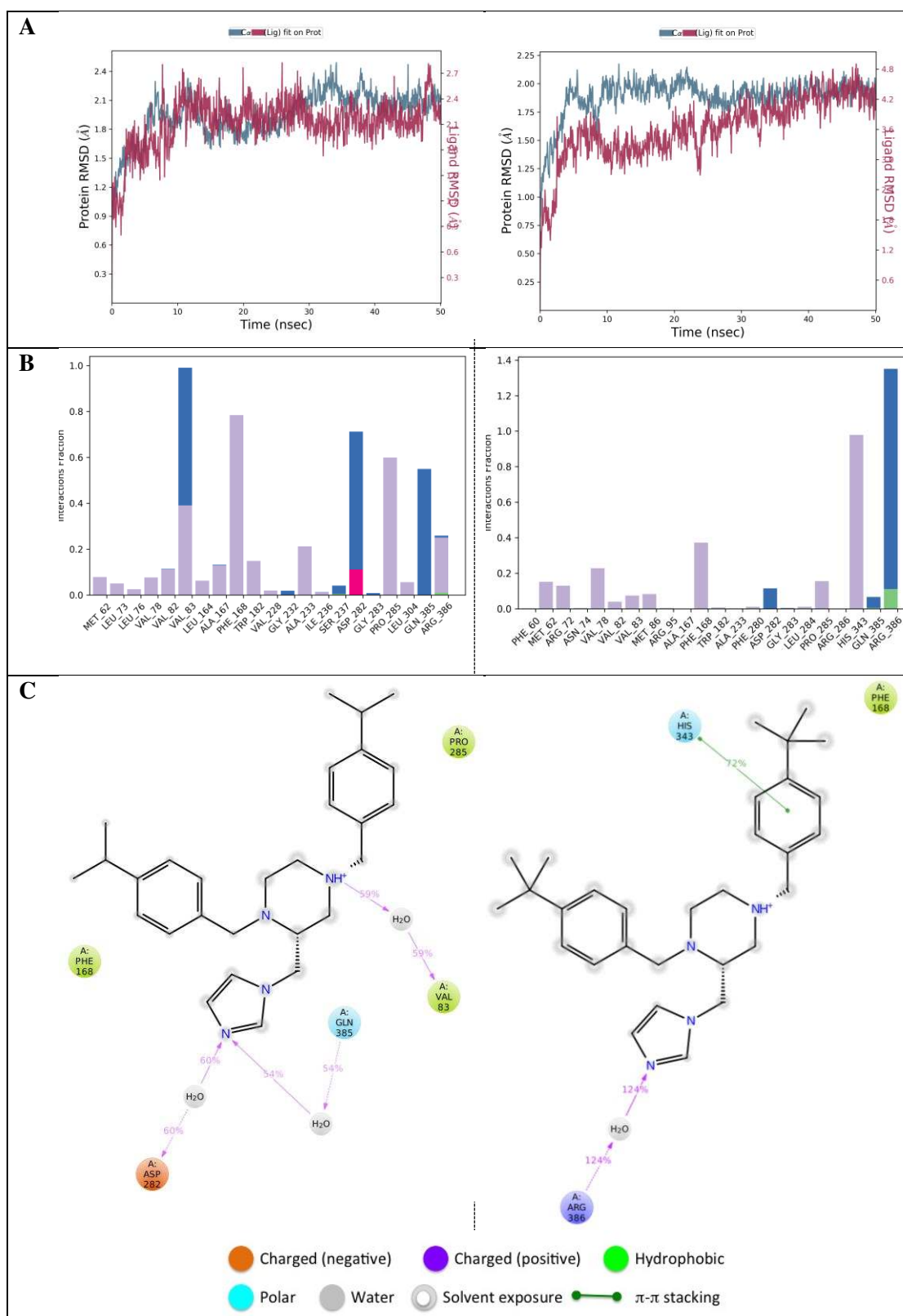


Figure 7. (A) Protein (blue) and ligand (red) root mean square of the CYP121A1-**8g** and **8h** complexes (B) Illustration of protein interactions with the ligand. The red column means ionic bond, green column a hydrogen bond, blue column water bridges, and a purple column a hydrophobic bond. The stacked bar charts are normalised over the course of the trajectory: for example, a value of 0.5 suggests that for 50% of the simulation time, the specific interaction is maintained (C) A schematic of detailed ligand atom interactions with the protein residues. Interactions that occur more than 30.0% of the simulation time in the selected trajectory (0.00 through 50.05 nsec), are shown (note: it is possible to have interactions with >100% as some residues may have multiple interactions of a single type with the same ligand atom e.g.

the ARG side chain has four H-bond donors that can all hydrogen-bond to a single H-bond acceptor).

Sterically, the transition from an isopropyl (**8g**) to a *tert*-butyl (**8h**) would appear sufficient to hinder closer binding interaction. As shown in Figure 8, the isopropyl derivative (**8g**) is able to take a position within the active site that is comparable (mode 1 binding) with the natural substrate cYY although one of the iPr-phenyl rings deviates from the cYY position through hydrophobic interaction with Pro285. However the *tert*-butyl derivative (**8h**) stabilises in the more open/flexible mode 2 conformation, with one of the tBu-phenyl rings deviating substantially, most likely owing to the reduction in conformational space within the binding pocket composed of Met62, Val82, Val83, Asn85 and Met86 (Figure 6).

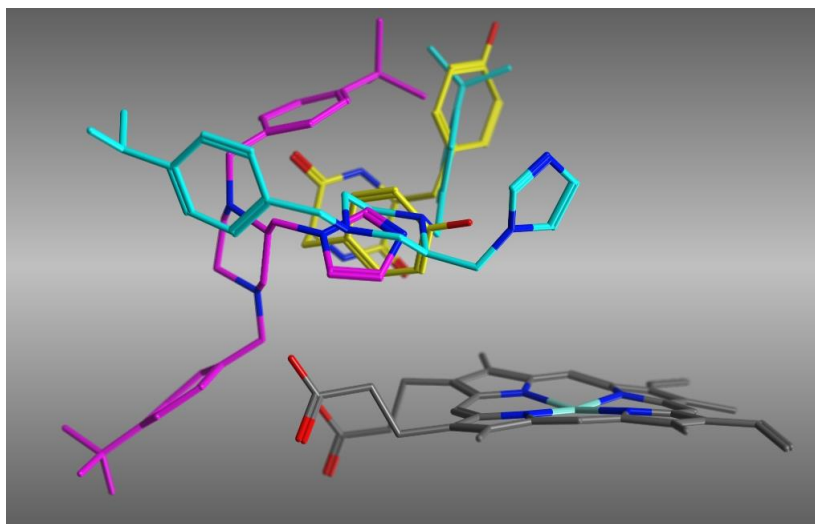


Figure 8. Positioning of the isopropyl derivative (**8g**, cyan) and *tert*-butyl derivative (**8h**, magenta) compared with cYY (yellow) after molecular dynamics simulation.

Both indirect binding modes 1 and 2 are possible and both would be effective in blocking the natural substrate cYY from binding at the active site. Crystallographic studies are needed to explore binding modes further.

3. Conclusions

With the exception of the pyridyl derivatives (**8i** and **8j**), all the new piperazine derivatives showed improved binding affinity to CYP121A1 and inhibitory activity against Mtb compared with the lead compound. Preliminary SAR for the described 1,4-dibenzyl-2-imidazol-1-yl-methylpiperazine derivatives would suggest that 4-substitution of the benzyl rings with either halides (**8a-8c**) or alkyl substituents (**8f-8h**) is beneficial for antimycobacterial activity with improved activity (MIC = 6.25 µg/mL) compared with clotrimazole (MIC = 20 µg/mL) (Table 1). Only one compound did not have substitution in the 4-position, namely the 3,5-dichloro derivative (**8d**), which had a reduced inhibitory activity compared with clotrimazole, however mono-substitutions in the 2- and 3-positions have not been investigated thus far and will be the focus of further studies to determine effect on binding and inhibitory activity. Two binding modes were observed from molecular modeling studies with the larger substituents (Br, 3,5-diCl, alkyl) showing a preference for the more open binding mode 2, while smaller substituents (F, Cl) showed a preference for the more compact binding mode 1 and more closely resemble the conformation of cYY (Table 2, Figure 6). The alkyl derivatives (**8f-8h**) were optimal with respect to binding affinity (K_D = 3.7, 1.6 and 1.2 µM, respectively), with the ethyl (**8f**) and isopropyl derivatives (**8g**) inducing the most extensive Soret absorbance shifts (416.5 to 422.5/423.5 nm) (Table 1). However, binding affinity did not clearly correlate with MIC, for example, the 4-chloro derivative (**8b**) and the *tert*-butyl derivative (**8g**) showed similar MIC values (6.25 µg/mL) but had completely different binding affinity to CYP121A1 (26.8 ± 7.9 compared with 1.6 ± 0.1 µM), suggesting different ligation modes with either indirect or direct haem coordination.

These piperazine derivatives are useful compounds for further development with future studies focusing on replacement of one of the phenyl rings to optimise positioning within the binding pocket composed of Met62, Val82, Val83, Asn85 and Met86 identified from the computational studies as restricting optimal fit within the CYP121A1 active site.

4. Experimental Section

4.1 General Experimental

All chemicals, reagents and solvents were purchased from Sigma-Aldrich, Fisher Scientific, Alfa-Aesar, Fluka and Acros Chemicals. Whenever required, solvents were dried prior to use as described by the handbook Purification of Laboratory Chemicals and stored over 4 Å molecular sieves under nitrogen. Flash column chromatography was performed with silica gel (230-400 mesh) (Merck) and TLC was performed on pre-coated silica gel plates (Merck Kiesel gel 60_{F254}, BDH). Melting points were determined on an electrothermal instrument (Gallenkamp), and are uncorrected. Compounds were visualised by irradiation with UV light at 254 nm and 365 nm or by using KMnO₄ stain or vanillin stain followed by heating. NMR spectra were recorded on a Bruker AVANCE DPX500 spectrometer operating at 500, 125 and 470 MHz for ¹H, ¹³C and ¹⁹F NMR, respectively, and auto calibrated to the deuterated solvent reference peak. The assignments were made using one-dimensional (1D) and two-dimensional (2D) HSQC and COSY spectra. Chemical shifts are given in δ relative to tetramethylsilane (TMS); the coupling constants (*J*) are given in Hertz. TMS was used as an internal standard (δ = 0 ppm) for ¹H NMR and CDCl₃ served as an internal standard (δ = 77.0 ppm) for ¹³C NMR. Multiplicity is denoted as s (singlet), br s (broad singlet), d (doublet), dd (doublet of doublet), t (triplet), q (quartet), m (multiplet) or

combinations thereof. High resolution mass spectra (HRMS) were determined at the EPSRC National Mass Spectrometry Facility at Swansea University and Medac Ltd., Brunel Science Centre, Surrey using ESI (Electrospray Ionisation). The following compounds were prepared as previously described: diimines **3a-3c**, **3e** and **3g-3j**;²⁹⁻³⁴ diamines **4a-4c**, **4e-4f** and **4i**;^{29,30,34,35} ethyl 1,4-bis(4-methoxybenzyl)piperazine-2-carboxylate (**5e**), (1,4-bis(4-methoxybenzyl)piperazin-2-yl)methanol (**6e**) and 2-(chloromethyl)-1,4-bis(4-methoxybenzyl)piperazine (**7e**).¹⁸ All compounds were more than 95% pure.

4.2 Chemistry

4.2.1. General method for the preparation of diimines (3). To a stirred solution of benzaldehyde (**1**) (2 m.eq.) in ethanol (1mL/mmol of benzaldehyde), was added ethylenediamine (**2**) (1 m.eq.) dropwise. The reaction mixture was stirred for 6 h at room temperature, and the precipitated white coloured crystals were collected by filtration, washed with diethyl ether and dried to give the desired diimine.

4.2.1.1. *N,N'*-(Ethane-1,2-diyl)bis(1-(3,5-dichlorophenyl)methanimine) (3d). Prepared from 3,5-dichlorobenzaldehyde (**1d**) (5 g, 28.6 mmol). Product obtained as a white solid, yield 5.10 g (96%). M.p. 130-132 °C. TLC (3:1 petroleum ether/EtOAc), *R_f* = 0.84. ¹H NMR (CDCl₃): δ 8.34 (s, 2H, 2 x CH=N), 7.74 (d, *J* = 1.9 Hz, 4H, Ar), 7.71 (t, *J* = 1.9 Hz, 2H, para-Ar), 3.91 (s, 4H, 2 x CH₂). ¹³C NMR (CDCl₃): δ 161.3 (2 x CH=N), 135.7 (2 x C, Ar), 135.3 (4 x C, Ar), 129.9 (2 x CH, para-Ar), 129.3 (4 x CH, Ar), 61.2 (2 x CH₂). [ESI-HRMS] calculated for C₁₆H₁₃Cl₄N₂: 372.9833 [M+H]⁺. Found: 372.9840 [M+H]⁺.

4.2.1.2. *N,N'*-(Ethane-1,2-diyl)bis(1-(4-ethylphenyl)methanimine) (3f). Prepared from 4-ethylbenzaldehyde (**1f**) (10 mL, 72.96 mmol). Product obtained as a white solid, yield 9.89 g (93%). M.p. 68-70 °C. TLC (4:1 petroleum ether/EtOAc), *R_f* = 0.84. ¹H NMR

(CDCl₃): δ 8.29 (s, 2H, 2 x CH=N), 7.62 (d, J = 8.1 Hz, 4H, Ar), 7.26 (d, J = 8.0 Hz, 4H, Ar), 3.84 (s, 4H, 2 x CH₂), 2.62 (q, J = 7.6 Hz, 4H, 2 x CH₂CH₃), 1.17 (t, J = 7.6 Hz, 6H, 2 x CH₂CH₃). ¹³C NMR (DMSO-d₆): δ 162.2 (2 x CH=N), 147.1 (2 x C, Ar), 134.2 (2 x C, Ar), 128.5 (4 x CH, Ar), 128.0 (4 x CH, Ar), 61.5 (2 x CH₂), 28.5 (2 x CH₂), 15.8 (2 x CH₃). [ESI-HRMS] calculated for C₂₀H₂₅N₂: 293.2018 [M+H]⁺. Found: 293.2033 [M+H]⁺.

4.2.2. General method for the preparation of diamines (4). To an ice-cooled stirred suspension of diimine (**3**) (1 m.eq.) in methanol (10 mL/mmol) was added sodium borohydride (4.6 m.eq.) in portions. The reaction mixture was stirred for 4 h at room temperature. After the reaction was complete, methanol was evaporated under vacuum and cold water (10 mL/mmol) was added to the remaining residue until cessation of effervescence. The mixture was extracted with EtOAc (10 mL/mmol), and the organic layer was washed with water (2 x 10 mL/mmol), brine (2 x 10 mL/mmol), dried (MgSO₄) and evaporated under vacuum.

4.2.2.1. *N*¹,*N*²-Bis(3,5-dichlorobenzyl)ethane-1,2-diamine (4d). Prepared from *N,N'*-(ethane-1,2-diyl)bis(1-(3,5-dichlorophenyl)methanimine) (**3d**) (5.0 g, 13.36 mmol). Product obtained as a white solid, yield 4.41 g (87%). M.p. 106-108 °C. TLC (2:1 petroleum ether/EtOAc), R_f = 0.12. ¹H NMR (CDCl₃): δ 7.28 (m, 2H, para-Ar), 7.26 (m, 4H, Ar), 3.77 (s, 4H, 2 x CH₂), 2.76 (s, 4H, 2 x CH₂), 1.79 (br s, 2H, 2 x NH). ¹³C NMR (CDCl₃): δ 143.6 (2 x C, Ar), 135.0 (4 x C, Ar), 127.2 (2 x CH, Ar), 126.5 (4 x CH, Ar), 52.8 (2 x CH₂), 48.4 (2 x CH₂). [ESI-HRMS] calculated for C₁₆H₁₇Cl₄N₂: 377.0146 [M+H]⁺. Found: 377.0159 [M+H]⁺.

4.2.2.2. *N*¹,*N*²-Bis(4-isopropylbenzyl)ethane-1,2-diamine (4g). Prepared from *N,N'*-(ethane-1,2-diyl)bis(1-(4-isopropylphenyl)methanimine) (**3g**) (10.4 g, 32.76 mmol). Product obtained as a white solid, yield 9.89 g (93%). M.p. 58-60 °C. TLC (2:1

petroleum ether/EtOAc), $R_f = 0.51$. ^1H NMR (CDCl_3): δ 7.22 (d, $J = 8.2$ Hz, 4H, Ar), 7.16 (d, $J = 7.8$ Hz, 4H, Ar), 3.62 (s, 4H, 2 x CH_2), 2.85 (m, 2H, 2 x $\text{CH}(\text{CH}_3)_2$), 2.57 (s, 4H, 2 x CH_2), 1.49 (br s, 2H, 2 x NH), 1.18 (d, $J = 6.8$ Hz, 12H, 4 x CH_3). ^{13}C NMR (DMSO-d_6): δ 146.9 (2 x C, Ar), 138.9 (2 x C, Ar), 128.3 (4 x CH, Ar), 126.4 (4 x CH, Ar), 53.3 (2 x CH_2), 48.9 (2 x CH_2), 33.6 (2 x $\text{CH}(\text{CH}_3)_2$), 24.43 (4 x CH_3). [ESI-HRMS] calculated for $\text{C}_{22}\text{H}_{33}\text{N}_2$: 325.2638 $[\text{M}+\text{H}]^+$. Found: 325.2643 $[\text{M}+\text{H}]^+$.

4.2.2.3. *N*¹,*N*²-Bis(4-(*tert*-butyl)benzyl)ethane-1,2-diamine (**4h**). Prepared from *N,N'*-(ethane-1,2-diyl)bis(1-(4-*tert*-butylphenyl)methanimine) (**3h**) (7.0 g, 20.08 mmol). Product obtained as a colourless oil, yield 6.5 g (92%). TLC (2:1 petroleum ether/EtOAc), $R_f = 0.53$. ^1H NMR (DMSO-d_6): δ 7.31 (d, $J = 10.0$ Hz, 4H, Ar), 7.21 (d, $J = 10.0$ Hz, 4H, Ar), 3.61 (s, 4H, 2 x CH_2), 2.56 (s, 4H, 2 x CH_2), 1.99 (br s, 2H, 2 x NH), 1.26 (s, 18H, 6 x CH_3). ^{13}C NMR (DMSO-d_6): δ 149.2 (2 x C, Ar), 138.3 (2 x C, Ar), 128.1 (4 x CH, Ar), 125.2 (4 x CH, Ar), 53.1 (2 x CH_2), 48.7 (2 x CH_2), 34.6 (2 x C), 31.7 (6 x CH_3). [ESI-HRMS] calculated for $\text{C}_{22}\text{H}_{37}\text{N}_2$: 353.2951 $[\text{M}+\text{H}]^+$. Found: 353.2948 $[\text{M}+\text{H}]^+$.

4.2.2.4. *N*¹,*N*²-Bis(pyridin-3-yl methyl)ethane-1,2-diamine (**4j**). Prepared from *N,N'*-(ethane-1,2-diyl)bis(1-(pyridine-3-yl)methanimine) (**3j**) (11.57 g, 48.57 mmol). Product obtained as a yellow oil, yield 3.7 g (31%). TLC (2:1 petroleum ether/EtOAc), $R_f = 0.12$. ^1H NMR (DMSO-d_6): δ 8.84 (s, 2H, pyridine), 8.60 (m, 2H, pyridine), 8.15 (t, $J = 5.8$ Hz, 2H, pyridine), 7.41 (d, $J = 5.7$ Hz, 2H, pyridine), 3.85 (s, 4H, 2 x CH_2), 3.71 (s, 4H, 2 x CH_2), 2.46 (br s, 2H, 2 x NH). ^{13}C NMR (DMSO-d_6): δ 150.3 (2 x CH, pyridine), 148.8 (CH, pyridine), 148.7 (CH, pyridine), 136.7 (2 x CH, pyridine), 134.7 (C, pyridine), 133.9 (C, pyridine), 123.7 (2 x CH, pyridine), 53.6 (2 x CH_2), 48.6 (2 x CH_2). [ESI-HRMS] calculated for $\text{C}_{14}\text{H}_{19}\text{N}_4$: 242.1531 $[\text{M}+\text{H}]^+$. Found: 242.1536 $[\text{M}+\text{H}]^+$.

4.2.3. General method for the preparation of ethyl 1,4-bis(aryl)piperazine-2-carboxylates (5). To a stirred solution of diamine (**4**) (1 m.eq., 2.84 g, 10.27 mmol) in anhydrous toluene at 80 °C (10 mL/mmol), was added triethylamine (2.5 m.eq.) and 2,3-dibromopropionic acid ethyl ester (1.05 m.eq.) dropwise, and the reaction mixture was heated at 80 °C overnight. The solvent was evaporated and the residue extracted with CH₂Cl₂ (10 mL/mmol), washed with saturated aqueous NaHCO₃ (5 x 5 mL/mmol) and brine (3 x 5 mL/mmol). The organic layer was dried (MgSO₄), concentrated under reduced pressure, and crude product was purified by gradient flash column chromatography.

4.2.3.1. Ethyl 1,4-bis(4-fluorobenzyl)piperazine-2-carboxylate (5a). Prepared from *N*¹,*N*²-bis(4-fluorobenzyl)ethane-1,2-diamine (**4a**) (2.84 g, 10.27 mmol). The product was eluted with petroleum ether – EtOAc 87.5:12.5 v/v to give the product as a white solid, yield 2.49 g (65%). M.p. 46-48 °C. TLC (4:1 petroleum ether/EtOAc), *R*_f = 0.82. ¹H NMR (CDCl₃): δ 7.29 (m, 4H, Ar), 6.99 (m, 4H, Ar), 4.19 (q, *J* = 7.0 Hz, 2H, CH₂CH₃), 3.89 (d, *J* = 13.3 Hz, 1H, H-7_a), 3.58 (d, *J* = 13.0 Hz, 1H, H-7_b), 3.53 (d, *J* = 13.2 Hz, 1H, H-8_a), 3.40 (d, *J* = 13.2 Hz, 1H, H-8_b), 3.30 (dd, *J* = 3.6, 5.5 Hz, 1H, H-2), 3.07 (m, 1H, H-3_a), 2.76 (m, 1H, H-3_b), 2.59-2.39 (m, 4H, H-6_{a,b}, H-5_{a,b}), 1.26 (t, *J* = 7.1 Hz, 3H, CH₂CH₃). ¹³C NMR (CDCl₃): δ 172.0 (C=O), 163.0 (C, Ar), 162.9 (C, Ar), 161.1 (C, Ar), 161.0 (C, Ar), 130.5 (2 x CH, Ar), 130.4 (2 x CH, Ar), 130.3 (2 x CH, Ar), 130.2 (2 x CH, Ar), 62.6 (CH, piperazine), 61.8 (CH₂CH₃), 60.4 (CH₂), 58.8 (CH₂), 55.4 (CH₂, piperazine), 53.0 (CH₂, piperazine), 48.5 (CH₂, piperazine), 14.3 (CH₂CH₃). ¹⁹F NMR (470 MHz, DMSO-*d*₆): δ -115.99 (2 x C₆H₅F). [ESI-HRMS] calculated for C₂₁H₂₅F₂N₂O₂: 375.1885 [M+H]⁺. Found: 375.1867 [M+H]⁺.

4.2.3.2. Ethyl 1,4-bis(4-chlorobenzyl)piperazine-2-carboxylate (5b). Prepared from *N*¹,*N*²-bis(4-chlorobenzyl)ethane-1,2-diamine (**4b**) (4.00 g, 12.93 mmol). The product

was eluted with petroleum ether – EtOAc 87.5:12.5 v/v to give the product as a white solid, yield 4.48 g (85%). M.p. 58-60 °C. TLC (4:1 petroleum ether/EtOAc), R_f = 0.60. ^1H NMR (CDCl_3): δ 7.26 (m, 8H, Ar), 4.19 (q, J = 7.1 Hz, 2H, CH_2CH_3), 3.90 (d, J = 13.5 Hz, 1H, H-7_a), 3.59 (d, J = 11.6 Hz, 1H, H-7_b), 3.54 (d, J = 13.4 Hz, 1H, H-8_a), 3.41 (d, J = 13.4 Hz, 1H, H-8_b), 3.32 (m, 1H, H-2), 3.08 (m, 1H, H-3_a), 2.77 (m, 1H, H-3_b), 2.59 (m, 1H, H-6_a), 2.53 (m, 1H, H-6_b), 2.40 (m, 2H, H-5_{a,b}), 1.26 (t, J = 7.1 Hz, 3H, CH_2CH_3). ^{13}C NMR (CDCl_3): δ 172.1 (C=O), 163.0 (2 x C, Ar), 161.1 (2 x C, Ar), 130.3 (4 x CH, Ar), 128.4 (4 x CH, Ar), 62.7 (CH, piperazine), 61.8 (CH_2CH_3), 60.4 (CH_2), 58.8 (CH_2), 55.4 (CH_2 , piperazine), 53.1 (CH_2 , piperazine), 48.4 (CH_2 , piperazine), 14.0 (CH_2CH_3). Anal. Calcd for $\text{C}_{21}\text{H}_{24}\text{Cl}_2\text{N}_2\text{O}_2$ (407.3335): C, 61.92; H, 5.94; N, 6.87. Found: C, 61.66; H, 5.91; N, 6.84.

4.2.3.3. *Ethyl 1,4-bis(4-bromobenzyl)piperazine-2-carboxylate (5c)*. Prepared from N^1,N^2 -bis(4-bromobenzyl)ethane-1,2-diamine (**4c**) (9.35 g, 23.48 mmol). The product was eluted with petroleum ether – EtOAc 87.5:12.5 v/v to give the product as a white solid, yield 9.68 g (83%). M.p. 64-66 °C. TLC (4:1 petroleum ether/EtOAc), R_f = 0.55. ^1H NMR (CDCl_3): δ 7.44 (m, 4H, Ar), 7.20 (m, 4H, Ar), 4.19 (q, J = 7.1 Hz, 2H, CH_2CH_3), 3.89 (d, J = 13.6 Hz, 1H, H-7_a), 3.59 (d, J = 13.4 Hz, 1H, H-7_b), 3.52 (d, J = 13.4 Hz, 1H, H-8_a), 3.39 (d, J = 13.4 Hz, 1H, H-8_b), 3.32 (dd, J = 3.4, 5.7 Hz, 1H, H-2), 3.08 (m, 1H, H-3_a), 2.79 (m, 1H, H-3_b), 2.55-2.38 (m, 4H, H-6_{a,b}, H-5_{a,b}), 1.26 (t, J = 7.1 Hz, 3H, CH_2CH_3). ^{13}C NMR (CDCl_3): δ 171.9 (C=O), 137.5 (2 x C, Ar), 137.1 (2 x C, Ar), 131.4 (2 x CH, Ar), 131.3 (2 x CH, Ar), 130.7 (2 x CH, Ar), 130.6 (2 x CH, Ar), 62.5 (CH, piperazine), 61.8 (CH_2CH_3), 60.5 (CH_2), 58.9 (CH_2), 55.4 (CH_2 , piperazine), 53.1 (CH_2 , piperazine), 48.4 (CH_2 , piperazine), 14.3 (CH_2CH_3). Anal. Calcd for $\text{C}_{21}\text{H}_{24}\text{Br}_2\text{N}_2\text{O}_2$ (496.2355): C, 50.83; H, 4.84; N, 5.64. Found: C, 50.51; H, 4.80; N, 5.57.

4.2.3.4. *Ethyl 1,4-bis(3,5-dichlorobenzyl)piperazine-2-carboxylate (5d)*. Prepared from *N*¹,*N*²-bis(3,5-dichlorobenzyl)ethane-1,2-diamine (**4d**) (6.0 g, 16.87 mmol). The product was eluted with petroleum ether – EtOAc 9:1 v/v to give the product as a buff solid, yield 4.46 g (71%). M.p. 80-84 °C. TLC (4:1 petroleum ether/EtOAc), *R*_f = 0.85. ¹H NMR (DMSO-*d*₆): δ 7.79 (m, 2H, para-Ar), 7.36 (m, 4H, Ar), 4.09 (q, *J* = 7.1 Hz, 2H, CH₂CH₃), 3.87 (d, *J* = 13.7 Hz, 1H, H-7_a), 3.72 (d, *J* = 14.0 Hz, 1H, H-7_b), 3.61 (d, *J* = 14.0 Hz, 1H, H-8_a), 3.47 (d, *J* = 14.0 Hz, 1H, H-8_b), 3.38 (m, 1H, H-2), 3.36 (m, 1H, H-3_a), 3.03 (m, 1H, H-3_b), 2.78 (m, 1H, H-6_a), 2.62 (m, 1H, H-6_b), 2.44-2.30 (m, 2H, H-5_{a,b}), 1.15 (t, *J* = 7.1 Hz, 3H, CH₂CH₃). ¹³C NMR (DMSO-*d*₆): δ 171.7 (C=O), 144.4 (2 x C, Ar), 143.4 (2 x C, Ar), 134.4 (C, Ar), 134.3 (C, Ar), 127.5 (2 x CH, Ar), 127.3 (2 x CH, Ar), 127.1 (CH, Ar), 126.9 (CH, Ar), 62.5 (CH, piperazine), 60.4 (CH₂CH₃), 60.4 (CH₂), 57.8 (CH₂), 55.2 (CH₂, piperazine), 53.3 (CH₂, piperazine), 48.5 (CH₂, piperazine), 14.5 (CH₂CH₃). Anal. Calcd for C₂₁H₂₂Cl₄N₂O₂ (476.2236): C, 52.96; H, 4.66; N, 5.88. Found: C, 52.69; H, 4.57; N, 5.81.

4.2.3.5. *Ethyl 1,4-bis(4-ethylbenzyl)piperazine-2-carboxylate (5f)*. Prepared from *N*¹,*N*²-bis(4-ethylbenzyl)ethane-1,2-diamine (**4f**) (9.0 g, 30.35 mmol). The product was eluted with petroleum ether – EtOAc 9:1 v/v to give the product as a colourless oil, yield 8.8 g (74%). TLC (95:5 CH₂Cl₂/MeOH), *R*_f = 0.97. ¹H NMR (DMSO-*d*₆): δ 7.15 (m, 8H, Ar), 4.06 (q, *J* = 7.7 Hz, 2H, CH₂CH₃), 3.79 (d, *J* = 13.4 Hz, 1H, H-7_a), 3.57 (d, *J* = 13.3 Hz, 1H, H-7_b), 3.48 (d, *J* = 13.2 Hz, 1H, H-8_a), 3.29 (m, 2H, H-8_b, H-2), 2.98 (m, 1H, H-3_a), 2.69 (m, 1H, H-3_b), 2.56 (q, *J* = 7.5 Hz, 4H, 2 x CH₂CH₃), 2.40 (m, 4H, H-6_{a,b}, H-5_{a,b}), 1.15 (m, 9H, 3 x CH₂CH₃). ¹³C NMR (DMSO-*d*₆): δ 171.7 (C=O), 142.8 (2 x C, Ar), 135.6 (2 x C, Ar), 129.0 (4 x CH, Ar), 128.0 (4 x CH, Ar), 64.7 (CH, piperazine), 63.0 (CH₂CH₃), 61.9 (CH₂), 60.7 (CH₂), 58.8 (CH₂, piperazine), 55.4 (CH₂, piperazine), 53.3 (CH₂, piperazine), 28.3 (2 x CH₂CH₃), 16.1 (2 x CH₂CH₃), 14.5

(CH₂CH₃). [ESI-HRMS] calculated for C₂₅H₃₅N₂O₂: 395.2699 [M+H]⁺. Found: 395.2692 [M+H]⁺.

4.2.3.6. *Ethyl 1,4-bis(4-isopropylbenzyl)piperazine-2-carboxylate (5g)*. Prepared from *N*¹,*N*²-bis(4-isopropylbenzyl)ethane-1,2-diamine (**4f**) (9.0 g, 27.73 mmol). The product was eluted with petroleum ether – EtOAc 9:1 v/v to give the product as a colourless oil, yield 10.0 g (85%). TLC (95:5 CH₂Cl₂/MeOH), R_f = 0.98. ¹H NMR (DMSO-d₆): δ 7.17 (m, 8H, Ar), 4.08 (q, *J* = 7.8 Hz, 2H, CH₂CH₃), 3.79 (d, *J* = 13.3 Hz, 1H, H-7_a), 3.57 (d, *J* = 13.2 Hz, 1H, H-7_b), 3.48 (d, *J* = 13.1 Hz, 1H, H-8_a), 3.27 (m, 2H, H-8_b, H-2), 2.99 (m, 1H, H-3_a), 2.84 (sept, 2H, *J* = 6.8 Hz, 2 x CH(CH₃)₂), 2.70 (m, 1H, H-3_b), 2.47-2.30 (m, 4H, H-6_{a,b}, H-5_{a,b}), 1.17 (d, *J* = 7.0 Hz, 12H, 4 x CH₃), 1.11 (t, *J* = 7.2 Hz, 3H, CH₂CH₃). ¹³C NMR (DMSO-d₆): δ 171.7 (C=O), 147.4 (2 x C, Ar), 136.3 (C, Ar), 135.8 (C, Ar), 129.0 (2 x CH, Ar), 128.9 (2 x CH, Ar), 126.5 (2 x CH, Ar), 126.4 (2 x CH, Ar), 65.4 (CH, piperazine), 61.9 (CH₂CH₃), 60.2 (CH₂), 60.1 (CH₂), 58.8 (CH₂, piperazine), 55.4 (CH₂, piperazine), 53.4 (CH₂, piperazine), 33.6 (2 x CH(CH₃)₂), 24.4 (4 x CH₃), 14.5 (CH₂CH₃). [ESI-HRMS] calculated for C₂₇H₃₉N₂O₂: 423.3012 [M+H]⁺. Found: 423.3016 [M+H]⁺.

4.2.3.7. *Ethyl 1,4-bis(4-(tert-butyl)benzyl)piperazine-2-carboxylate (5h)*. Prepared from *N*¹,*N*²-bis(4-*tert*-butylbenzyl)ethane-1,2-diamine (**4h**) (6.0 g, 17.01 mmol). The product was eluted with petroleum ether – EtOAc 92.5:7.5 v/v to give the product as a white solid, yield 5.0 g (65%). M.p. 82-84 °C. TLC (5:1 petroleum ether/EtOAc), R_f = 0.83. ¹H NMR (CDCl₃): δ 7.31 (m, 4H, Ar), 7.18 (m, 4H, Ar), 4.06 (q, *J* = 7.1 Hz, 2H, CH₂CH₃), 3.80 (d, *J* = 13.1 Hz, 1H, H-7_a), 3.54 (d, *J* = 13.2 Hz, 1H, H-7_b), 3.39 (m, 2H, H-8_{a,b}), 3.30 (dd, *J* = 3.6, 5.6 Hz, 1H, H-2), 2.99 (m, 1H, H-3_a), 2.72 (m, 1H, H-3_b), 2.41 (m, 4H, H-6_{a,b}, H-5_{a,b}), 1.25 (s, 18H, 6 x CH₃), 1.11 (t, *J* = 6.7 Hz, 3H, CH₂CH₃). ¹³C NMR (CDCl₃): δ 171.5 (C=O), 135.8 (2 x C, Ar), 132.2 (2 x C, Ar), 128.8 (4 x CH,

Ar), 125.4 (4 x CH, Ar), 64.1 (CH, piperazine), 63.0 (CH₂CH₃), 61.8 (CH₂), 60.3 (CH₂), 58.6 (CH₂, piperazine), 53.2 (CH₂, piperazine), 47.9 (CH₂, piperazine), 34.6 (2 x C(CH₃)₃), 31.6 (6 x CH₃), 14.5 (CH₂CH₃). [ESI-HRMS] calculated for C₂₉H₄₃N₂O₂: 451.3324 [M+H]⁺. Found: 451.3327 [M+H]⁺.

4.2.3.8. *Ethyl 1,4-bis(pyridin-4-ylmethyl)piperazine-2-carboxylate (5i)*. Prepared from *N*¹,*N*²-bis(pyridin-4-yl)ethane-1,2-diamine (**4i**) (3.66 g, 15.10 mmol). The product was eluted with CH₂Cl₂-MeOH-Et₃N 97.5:1.5:1 v/v/v to give the product as a brown oil, yield 3.36 g (65%). TLC (95:5 CH₂Cl₂/MeOH), R_f = 0.32. ¹H NMR (DMSO-*d*₆): δ 8.50 (m, 4H, Ar), 7.30 (m, 4H, Ar), 4.10 (q, *J* = 7.6 Hz, 2H, CH₂CH₃), 3.90 (d, *J* = 13.0 Hz, 1H, H-7_a), 3.74 (d, *J* = 13.7 Hz, 1H, H-7_b), 3.61 (d, *J* = 13.4 Hz, 1H, H-8_a), 3.46 (d, *J* = 13.4 Hz, 1H, H-8_b), 3.40 (dd, *J* = 3.6, 5.9 Hz, 1H, H-2), 3.04 (m, 1H, H-3_a), 2.81 (m, 1H, H-3_b), 2.58 (m, 1H, H-6_a), 2.45 (m, 1H, H-6_b), 2.36 (m, 2H, H-5_{a,b}), 1.15 (t, *J* = 7.0 Hz, 3H, CH₂CH₃). ¹³C NMR (DMSO-*d*₆): δ 171.6 (C=O), 150.0 (4 x CH, Ar), 148.7 (C, Ar), 147.7 (C, Ar), 124.0 (2 x CH, Ar), 123.8 (2 x CH, Ar), 62.0 (CH, piperazine), 60.7 (CH₂CH₃), 60.4 (CH₂), 57.8 (CH₂), 55.4 (CH₂, piperazine), 53.4 (CH₂, piperazine), 52.5 (CH₂, piperazine), 14.6 (CH₂CH₃). [ESI-HRMS] calculated for C₁₉H₂₅N₄O₂: 341.1899 [M+H]⁺. Found: 341.1910 [M+H]⁺.

4.2.3.9. *Ethyl 1,4-bis(pyridin-3-ylmethyl)piperazine-2-carboxylate (5j)*. Prepared from *N*¹,*N*²-bis(pyridin-3-yl)ethane-1,2-diamine (**4j**) (3.50 g, 14.44 mmol). The product was eluted with CH₂Cl₂-MeOH-Et₃N 96.5:2.5:1 v/v/v to give the product as a brown oil, yield 2.30 g (47%). TLC (95:5 CH₂Cl₂/MeOH), R_f = 0.31. ¹H NMR (DMSO-*d*₆): δ 8.47 (s, 2H, pyridine), 8.46 (m, 2H, pyridine), 7.67 (t, *J* = 7.7 Hz, 2H, pyridine), 7.35 (d, *J* = 5.6 Hz, 2H, pyridine), 4.08 (q, *J* = 7.8 Hz, 2H, CH₂CH₃), 3.88 (d, *J* = 14.0 Hz, 1H, H-7_a), 3.70 (d, *J* = 13.8 Hz, 1H, H-7_b), 3.60 (d, *J* = 13.5 Hz, 1H, H-8_a), 3.38 (m, 2H, H-8_b, H-2), 3.00 (m, 1H, H-3_a), 2.75 (m, 1H, H-3_b), 2.54 (m, 1H, H-6_a), 2.42 (m, 1H, H-

6b), 2.34 (m, 2H, H-5_{a,b}), 1.13 (t, $J = 7.7$ Hz, 3H, CH₂CH₃). ¹³C NMR (DMSO-d₆): δ 171.6 (C=O), 150.4 (CH, pyridine), 150.2 (CH, pyridine), 148.8 (CH, pyridine), 148.7 (CH, pyridine), 136.9 (CH, pyridine), 136.6 (CH, pyridine), 134.7 (C, pyridine), 133.9 (C, pyridine), 123.9 (CH, pyridine), 123.8 (CH, pyridine), 62.3 (CH, piperazine), 60.4 (CH₂CH₃), 59.1 (CH₂), 56.3 (CH₂), 56.0 (CH₂, piperazine), 55.2 (CH₂, piperazine), 53.2 (CH₂, piperazine), 14.5 (CH₂CH₃). [ESI-HRMS] calculated for C₁₉H₂₅N₄O₂: 341.1899 [M+H]⁺. Found: 341.1903 [M+H]⁺.

4.2.4. General method for the preparation of alcohols (6). To an ice-cooled solution of ethyl carboxylate (**5**) (1 m.eq.) in dry THF (3 mL/mmol) was added LiAlH₄ (1M in THF, 1.5 m.eq.) dropwise over 25 min. The reaction was then stirred at room temperature overnight, then cooled in an ice-bath and carefully quenched with H₂O until cessation of effervescence. The reaction mixture was extracted with EtOAc (2 x 10 mL/mmol), then the combined organic layers washed with H₂O (3 x 10 mL/mmol), dried (MgSO₄) and concentrated under reduced pressure. The crude product was purified by gradient flash column chromatography.

4.2.4.1. (1,4-Bis(4-fluorobenzyl)piperazin-2-yl)methanol (6a). Prepared from ethyl 1,4-bis(4-fluorobenzyl)piperazine-2-carboxylate (**5a**) (1.2 g, 3.20 mmol). The product was eluted with CH₂Cl₂-MeOH 95:5 v/v to give the product as a yellow oil, yield 0.86 g (81%). TLC (95:5 CH₂Cl₂/MeOH), $R_f = 0.55$. ¹H NMR (CDCl₃): δ 7.30 (m, 8H, Ar), 3.61 (m, 2H, H-9_{a,b}), 3.48 (m, 2H, H-7_{a,b}), 2.96 (m, 2H, H-8_{a,b}), 2.67 (m, 3H, H-2, H-3_{a,b}), 2.59 (m, 1H, H-6_a), 2.53-2.45 (m, 2H, H-6_b, H-5_a), 2.37 (m, 1H, H-5_b). ¹³C NMR (CDCl₃): δ 163.1 (C, Ar), 161.1 (C, Ar), 138.4 (C, Ar), 137.5 (C, Ar), 128.9 (4 x CH, Ar), 128.4 (4 x CH, Ar), 62.3 (CH₂), 58.7 (CH, piperazine), 58.0 (CH₂), 57.2 (CH₂), 55.9 (CH₂, piperazine), 52.4 (CH₂, piperazine), 49.9 (CH₂, piperazine). ¹⁹F NMR

(CDCl₃): δ -116.00 (2 x C₆H₅F). [ESI-HRMS] calculated for C₁₉H₂₃F₂N₂O: 333.1807 [M+H]⁺. Found: 333.1823 [M+H]⁺.

4.2.4.2. (1,4-Bis(4-chlorobenzyl)piperazin-2-yl)methanol (**6b**). Prepared from ethyl 1,4-bis(4-chlorobenzyl)piperazine-2-carboxylate (**5b**) (1.0 g, 2.45 mmol). The product was eluted with CH₂Cl₂-MeOH 97:3 v/v to give the product as a colourless oil, yield 0.52 g (59%). TLC (95:5 CH₂Cl₂/MeOH), R_f = 0.59. ¹H NMR (DMSO-d₆): δ 7.30 (m, 8H, Ar), 4.50 (t, *J* = 5.1 Hz, 1H, OH), 3.69 (m, 2H, H-9_{a,b}), 3.43 (m, 2H, H-7_{a,b}), 3.38 (d, *J* = 13.6 Hz, 1H, H-8_a), 3.27 (d, *J* = 13.6 Hz, 1H, H-8_b), 2.73 (m, 1H, H-2), 2.59 (m, 1H, H-3_a), 2.48 (m, 1H, H-3_b), 2.41 (m, 1H, H-6_a), 2.18-2.02 (m, 3H, H-6_b, H-5_{a,b}). ¹³C NMR (DMSO-d₆): δ 138.9 (C, Ar), 137.8 (C, Ar), 131.9 (C, Ar), 131.6 (C, Ar), 130.9 (2 x CH, Ar), 130.8 (2 x CH, Ar), 128.6 (2 x CH, Ar), 128.5 (2 x CH, Ar), 62.7 (CH₂), 61.8 (CH, piperazine), 61.6 (CH₂), 58.2 (CH₂), 57.3 (CH₂, piperazine), 52.9 (CH₂, piperazine), 51.1 (CH₂, piperazine). [ESI-HRMS] calculated for C₁₉H₂₃Cl₂N₂O: 365.1187 [M+H]⁺. Found: 365.1190 [M+H]⁺.

4.2.4.3. (1,4-Bis(4-bromobenzyl)piperazin-2-yl)methanol (**6c**). Prepared from ethyl 1,4-bis(4-bromobenzyl)piperazine-2-carboxylate (**5c**) (1.20 g, 2.41 mmol). The product was eluted with CH₂Cl₂-MeOH 97.5:2.5 v/v to give the product as a white solid, yield 0.89 g (81%). M.p. 88-90 °C. TLC (95:5 CH₂Cl₂/MeOH), R_f = 0.63. ¹H NMR (DMSO-d₆): δ 7.49 (m, 4H, Ar), 7.25 (m, 4H, Ar), 4.54 (t, *J* = 5.1 Hz, 1H, OH), 3.66 (m, 2H, H-9_{a,b}), 3.42 (m, 2H, H-7_{a,b}), 3.38 (d, *J* = 13.8 Hz, 1H, H-8_a), 3.26 (d, *J* = 13.9 Hz, 1H, H-8_b), 2.69 (m, 1H, H-2), 2.56 (m, 1H, H-3_a), 2.47 (m, 1H, H-3_b), 2.40 (m, 1H, H-6_a), 2.17-2.02 (m, 3H, H-6_b, H-5_{a,b}). ¹³C NMR (DMSO-d₆): δ 138.2 (2 x C, Ar), 131.5 (4 x CH, Ar), 131.2 (4 x CH, Ar), 120.3 (2 x C, Ar), 62.7 (CH₂), 61.8 (CH, piperazine), 61.6 (CH₂), 58.1 (CH₂), 56.4 (CH₂, piperazine), 53.0 (CH₂, piperazine), 51.1 (CH₂,

piperazine). [ESI-HRMS] calculated for $C_{19}H_{23}Br_2N_2O$: 453.0177 $[M+H]^+$. Found: 453.0163 $[M+H]^+$.

4.2.4.4. (1,4-Bis(3,5-dichlorobenzyl)piperazin-2-yl)methanol (**6d**). Prepared from ethyl 1,4-bis(3,5-dichlorobenzyl)piperazine-2-carboxylate (**5d**) (1.5 g, 3.14 mmol). The product was eluted with petroleum ether – EtOAc 7:3 v/v v/v to give the product as a yellow oil, yield 0.75 g (56%). TLC (1:1 petroleum ether/EtOAc), R_f = 0.31. 1H NMR (DMSO- d_6): δ 7.46 (t, J = 1.7 Hz, 1H, para-Ar), 7.43 (t, J = 1.7 Hz, 1H, para-Ar), 7.36 (d, J = 1.7 Hz, 2H, Ar), 7.33 (d, J = 1.7 Hz, 2H, Ar), 4.60 (t, J = 5.1 Hz, 1H, OH), 3.63 (m, 2H, H-9_{a,b}), 3.45 (m, 2H, H-7_{a,b}), 3.37 (m, 2H, H-8_{a,b}), 2.68 (m, 1H, H-2), 2.58 (m, 1H, H-3_a), 2.48 (m, 1H, H-3_b), 2.43 (m, 1H, H-6_a), 2.21-2.07 (m, 3H, H-6_b, H-5_{a,b}). ^{13}C NMR (DMSO- d_6): δ 142.5 (2 x C, Ar), 141.3 (2 x C, Ar), 133.4 (2 x C, Ar), 130.5 (CH, Ar), 130.4 (CH, Ar), 128.6 (CH, Ar), 128.5 (CH, Ar), 127.9 (CH, Ar), 127.7 (CH, Ar), 61.7 (CH₂), 61.6 (CH, piperazine), 61.5 (CH₂), 57.4 (CH₂), 56.9 (CH₂, piperazine), 52.8 (CH₂, piperazine), 51.1 (CH₂, piperazine). [ESI-HRMS] calculated for $C_{19}H_{21}Cl_4N_2O$: 433.0408 $[M+H]^+$. Found: 433.0399 $[M+H]^+$.

4.2.4.5. (1,4-Bis(4-ethylbenzyl)piperazin-2-yl)methanol (**6f**). Prepared from ethyl 1,4-bis(4-ethylbenzyl)piperazine-2-carboxylate (**5f**) (2.0 g, 5.06 mmol). The product was eluted with CH₂Cl₂-MeOH 97:3 v/v to give the product as a buff solid, yield 1.48 g (83%). M.p. 40-42 °C. TLC (95:5 CH₂Cl₂/MeOH), R_f = 0.65. 1H NMR (DMSO- d_6): δ 7.18 (m, 8H, Ar), 4.52 (br s, 1H, OH), 3.70 (m, 2H, H-9_{a,b}), 3.38 (m, 2H, H-7_{a,b}), 3.22 (d, J = 13.7 Hz, 1H, H-8_a), 2.73 (d, J = 13.4 Hz, 1H, H-8_b), 2.57 (q, J = 7.4 Hz, 4H, 2 x CH₂CH₃), 2.38 (m, 1H, H-2), 2.14-2.02 (m, 6H, H-3_{a,b}, H-6_{a,b}, H-5_{a,b}), 1.16 (t, J = 7.4 Hz, 6H, 2 x CH₃). ^{13}C NMR (DMSO- d_6): δ 142.7 (C, Ar), 142.4 (C, Ar), 136.8 (C, Ar), 135.9 (C, Ar), 129.2 (2 x CH, Ar), 129.1 (2 x CH, Ar), 128.0 (2 x CH, Ar), 127.9 (2 x CH, Ar), 62.5 (CH₂), 61.7 (CH, piperazine), 60.2 (CH₂), 57.7 (CH₂), 56.4 (CH₂,

piperazine), 53.0 (CH₂, piperazine), 51.1 (CH₂, piperazine), 28.3 (2 x CH₂CH₃), 16.1 (2 x CH₂CH₃). [ESI-HRMS] calculated for C₂₃H₃₃N₂O: 353.2587 [M+H]⁺. Found: 353.2610 [M+H]⁺.

4.2.4.6. (1,4-Bis(4-isopropylbenzyl)piperazin-2-yl)methanol (**6g**). Prepared from ethyl 1,4-bis(4-isopropylbenzyl)piperazine-2-carboxylate (**5g**) (2.50 g, 5.91 mmol). The product was eluted with CH₂Cl₂-MeOH 97:3 v/v to give the product as a buff solid, yield 1.91 g (85%). M.p. 68-70 °C. TLC (95:5 CH₂Cl₂/MeOH), R_f = 0.68. ¹H NMR (DMSO-d₆): δ 7.17 (m, 8H, Ar), 4.53 (br s, 1H, OH), 3.71 (m, 2H, H-9_{a,b}), 3.40 (d, *J* = 13.1 Hz, 1H, H-7_a), 3.37 (d, *J* = 13.1 Hz, 1H, H-7_b), 3.21 (d, *J* = 13.8 Hz, 1H, H-8_a), 2.85 (sept, *J* = 3.2 Hz, 2H, 2 x CH(CH₃)₂), 2.74 (d, *J* = 13.2 Hz, 1H, H-8_b), 2.58 (m, 1H, H-2), 2.47-2.39 (m, 2H, H-3_{a,b}), 2.15-2.01 (m, 4H, H-6_{a,b}, H-5_{a,b}), 1.19 (d, *J* = 1.9 Hz, 6H, 2 x CH₃), 1.17 (d, *J* = 1.8 Hz, 6H, 2 x CH₃). ¹³C NMR (DMSO-d₆): δ 137.0 (2 x C, Ar), 136.0 (2 x C, Ar), 129.2 (2 x CH, Ar), 129.0 (2 x CH, Ar), 126.5 (2 x CH, Ar), 126.4 (2 x CH, Ar), 62.5 (CH₂), 61.8 (CH, piperazine), 60.2 (CH₂), 57.9 (CH₂), 56.5 (CH₂, piperazine), 53.1 (CH₂, piperazine), 51.2 (CH₂, piperazine), 33.6 (2 x CH(CH₃)₂), 24.4 (4 x CH₃). [ESI-HRMS] calculated for C₂₅H₃₇N₂O: 381.2900 [M+H]⁺. Found: 381.2890 [M+H]⁺.

4.2.4.7. (1,4-Bis(4-(tert-butyl)benzyl)piperazin-2-yl)methanol (**6h**). Prepared from ethyl 1,4-bis(4-tert-butylbenzyl)piperazine-2-carboxylate (**5h**) (5 g, 11.09 mmol). The product was eluted with CH₂Cl₂-MeOH 97:3 v/v to give the product as a white solid, yield 4.12 g (91%). M.p. 92-94 °C. TLC (95:5 CH₂Cl₂/MeOH), R_f = 0.71. ¹H NMR (DMSO-d₆): δ 7.30 (m, 4H, Ar), 7.21 (m, 4H, Ar), 4.55 (br s, 1H, OH), 3.70 (m, 2H, H-9_{a,b}), 3.41 (d, *J* = 13.2 Hz, 1H, H-7_a), 3.23 (d, *J* = 13.1 Hz, 1H, H-7_b), 2.77 (d, *J* = 12.6 Hz, 1H, H-8_a), 2.58 (d, *J* = 12.7 Hz, 1H, H-8_b), 2.44 (m, 3H, H-2, H-3_{a,b}), 2.41 (m, 2H, H-6_{a,b}), 2.06 (m, 2H, H-5_{a,b}), 1.27 (s, 9H, 3 x CH₃), 1.26 (s, 9H, 3 x CH₃). ¹³C NMR

(DMSO- d_6): δ 149.5 (C, Ar), 149.2 (C, Ar), 136.6 (C, Ar), 135.6 (C, Ar), 128.9 (2 x CH, Ar), 128.8 (2 x CH, Ar), 125.2 (2 x CH, Ar), 125.1 (2 x CH, Ar), 62.5 (CH₂), 61.8 (CH, piperazine), 60.2 (CH₂), 57.9 (CH₂), 56.6 (CH₂, piperazine), 53.1 (CH₂, piperazine), 51.2 (CH₂, piperazine), 34.5 (2 x C(CH₃)₃), 31.6 (6 x CH₃). [ESI-HRMS] calculated for C₂₇H₄₁N₂O: 409.3213 [M+H]⁺. Found: 409.3215 [M+H]⁺.

4.2.4.8. *(1,4-Bis(pyridine-4-yl)piperazin-2-yl)methanol (6i)*. Prepared from ethyl 1,4-bis(pyridin-4-yl)piperazine-2-carboxylate (**5i**) (0.96 g, 2.82 mmol). The product was eluted with CH₂Cl₂-MeOH-Et₃N 89:10:1 v/v/v to give the product as a brown oil, yield 0.60 g (71%). TLC (95:5 CH₂Cl₂/MeOH), R_f = 0.25. ¹H NMR (DMSO- d_6): δ 8.49 (m, 4H, Ar), 7.32 (m, 4H, Ar), 4.58 (t, J = 5.2 Hz, 1H, OH), 3.65 (m, 2H, H-9_{a,b}), 3.49 (m, 2H, H-7_{a,b}), 3.39 (m, 2H, H-8_{a,b}), 2.70 (m, 1H, H-2), 2.61 (m, 1H, H-3_a), 2.46 (m, 1H, H-3_b), 2.26-2.16 (m, 3H, H-6_{a,b}, H-5_a), 2.10 (m, 1H, H-5_b). ¹³C NMR (DMSO- d_6): δ 136.8 (2 x C, pyridine), 129.3 (4 x CH, pyridine), 126.7 (4 x CH, pyridine), 62.5 (CH₂), 61.6 (CH, piperazine), 60.1 (CH₂), 57.5 (CH₂), 56.4 (CH₂, piperazine), 52.9 (CH₂, piperazine), 51.4 (CH₂, piperazine). [ESI-HRMS] calculated for C₁₇H₂₃N₄O: 299.1872 [M+H]⁺. Found: 299.1882 [M+H]⁺.

4.2.4.9. *(1,4-Bis(pyridine-3-yl)piperazin-2-yl)methanol (6j)*. Prepared from ethyl 1,4-bis(pyridin-3-yl)piperazine-2-carboxylate (**5j**) (1.10 g, 3.23 mmol). The product was eluted with CH₂Cl₂-MeOH-Et₃N 89:10:1 v/v/v to give the product as a brown oil, yield 0.68 g (71%). TLC (95:5 CH₂Cl₂/MeOH), R_f = 0.24. ¹H NMR (DMSO- d_6): δ 8.49 (s, 1H, pyridine), 8.47 (s, 1H, pyridine), 8.45 (m, 2H, pyridine), 7.70 (t, J = 7.7 Hz, 2H, pyridine), 7.34 (d, J = 5.6 Hz, 2H, pyridine), 4.57 (t, J = 5.2 Hz, 1H, OH), 3.70 (m, 2H, H-9_{a,b}), 3.48 (m, 2H, H-7_{a,b}), 3.41 (m, 2H, H-8_{a,b}), 2.70 (m, 1H, H-2), 2.57 (m, 1H, H-3_a), 2.41 (m, 1H, H-3_b), 2.20-2.04 (m, 4H, H-6_{a,b}, H-5_{a,b}). ¹³C NMR (DMSO- d_6): δ 150.1 (CH, pyridine), 149.8 (CH, pyridine), 148.5 (CH, pyridine), 147.6 (CH, pyridine),

136.8 (2 x CH, pyridine), 134.3 (2 x C, pyridine), 121.9 (2 x CH, pyridine), 62.4 (CH₂), 61.2 (CH, piperazine), 59.7 (CH₂), 56.3 (CH₂), 53.6 (CH₂, piperazine), 51.3 (CH₂, piperazine), 49.8 (CH₂, piperazine). [ESI-HRMS] calculated for C₁₇H₂₃N₄O: 299.1872 [M+H]⁺. Found: 299.1879 [M+H]⁺.

4.2.5. General method for the preparation of chlorides (7). To an ice-cooled solution of alcohol (**6**) (1 m.eq.) in dry CH₂Cl₂ (5 mL/mmol) was added thionyl chloride (10 m.eq.) dropwise over 25 min. The reaction was stirred at room temperature for 48 h then cooled in an ice-bath and carefully quenched with saturated aqueous NaHCO₃ in portions until slightly basic (pH 8.0). The organic layer was separated, washed with brine (3 x 10 mL/mmol), H₂O (2 x 10 mL/mmol), dried (MgSO₄) and evaporated under reduced pressure to give the chloride. Chloride products (**7a-7d**) that were highly sensitive to light and moisture were wrapped with aluminium foil, stored in the freezer immediately once prepared, and used in the next step without further purification. Stable chloride products (**7f-7j**) were purified by gradient column chromatography.

4.2.5.1. 2-(Chloromethyl)-1,4-bis(4-fluorobenzyl)piperazine (7a). Prepared from (1,4-bis(4-fluorobenzyl)piperazin-2-yl)methanol (**6a**) (1.00 g, 3.00 mmol). The product was obtained as a brown solid, yield 0.52 g (50%). M.p. 66-68 °C. TLC (95:5 CH₂Cl₂/MeOH), R_f = 0.91. ¹H NMR (DMSO-d₆): δ 7.36-7.12 (m, 8H, Ar), 3.93 (m, 2H, H-9_{a,b}), 3.84 (d, *J* = 13.2 Hz, 1H, H-7_a), 3.49 (d, *J* = 13.3 Hz, 1H, H-7_b), 3.41 (m, 2H, H-8_{a,b}), 2.71 (m, 1H, H-2), 2.61 (m, 1H, H-3_a), 2.48 (m, 1H, H-3_b), 2.31 (m, 4H, H-6_{a,b}, H-5_{a,b}). ¹³C NMR (DMSO-d₆): δ 162.7 (2 x C, Ar), 139.3 (2 x C, Ar), 129.1 (2 x CH, Ar), 128.7 (2 x CH, Ar), 115.5 (2 x CH, Ar), 115.3 (2 x CH, Ar), 61.5 (CH₂), 59.7 (CH, piperazine), 57.5 (CH₂), 54.9 (CH₂, piperazine), 54.7 (CH₂, piperazine), 49.1 (CH₂, piperazine), 43.4 (CH₂). ¹⁹F NMR (DMSO-d₆): δ -115.99 (2 x C₆H₅F). [ESI-HRMS] calculated for C₁₉H₂₂ClF₂N₂: 351.1440 [M+H]⁺. Found: 351.1449 [M+H]⁺.

4.2.5.2. 2-(Chloromethyl)-1,4-bis(4-chlorobenzyl)piperazine (**7b**). Prepared from (1,4-bis(4-chlorobenzyl)piperazin-2-yl)methanol (**6b**) (0.3 g, 0.82 mmol). The product was obtained as a brown solid, yield 0.21 g (68%). M.p. 64-66 °C. TLC (95:5 CH₂Cl₂/MeOH), R_f = 0.92. ¹H NMR (DMSO-d₆): δ 7.33 (m, 8H, Ar), 3.95 (m, 2H, H-9_{a,b}), 3.48-3.36 (m, 4H, H-7_{a,b}, H-8_{a,b}), 2.72 (m, 1H, H-2), 2.60 (m, 1H, H-3_a), 2.49 (m, 1H, H-3_b), 2.38 (m, 2H, H-6_{a,b}), 2.28 (m, 2H, H-5_{a,b}). ¹³C NMR (DMSO-d₆): δ 139.1 (2 x C, Ar), 131.8 (2 x C, Ar), 130.8 (2 x CH, Ar), 129.1 (2 x CH, Ar), 128.6 (2 x CH, Ar), 127.3 (2 x CH, Ar), 62.4 (CH₂), 59.7 (CH, piperazine), 58.1 (CH₂), 57.5 (CH₂, piperazine), 52.8 (CH₂, piperazine), 49.2 (CH₂, piperazine), 43.3 (CH₂). [ESI-HRMS] calculated for C₁₉H₂₂Cl₃N₂: 383.0849 [M+H]⁺. Found: 383.0856 [M+H]⁺.

4.2.5.3. 2-(Chloromethyl)-1,4-bis(4-bromobenzyl)piperazine (**7c**). Prepared from (1,4-bis(4-bromobenzyl)piperazin-2-yl)methanol (**6c**) (0.80 g, 1.76 mmol). The product was obtained as a light yellow solid, yield 0.53 g (64%). M.p. 70-72 °C. TLC (95:5 CH₂Cl₂/MeOH), R_f = 0.94. ¹H NMR (DMSO-d₆): δ 7.50 (d, *J* = 8.3 Hz, 4H, Ar), 7.27 (d, *J* = 8.3 Hz, 4H, Ar), 3.98 (m, 2H, H-9_{a,b}), 3.85 (m, 2H, H-7_{a,b}), 3.45 (m, 2H, H-8_{a,b}), 3.40 (m, 1H, H-2), 2.70 (m, 1H, H-3_a), 2.60 (m, 1H, H-3_b), 2.45 (m, 1H, H-6_a), 2.37 (m, 1H, H-6_b), 2.26 (m, 2H, H-5_{a,b}). ¹³C NMR (DMSO-d₆): δ 137.7 (2 x C, Ar), 137.2 (2 x C, Ar), 131.4 (2 x CH, Ar), 131.3 (2 x CH, Ar), 130.6 (2 x CH, Ar), 130.4 (2 x CH, Ar), 62.1 (CH₂), 60.3 (CH, piperazine), 57.5 (CH₂), 55.1 (CH₂, piperazine), 52.7 (CH₂, piperazine), 51.0 (CH₂, piperazine), 42.3 (CH₂). [ESI-HRMS] calculated for C₁₉H₂₂Br₂ClN₂: 470.9838 [M+H]⁺. Found: 470.9844 [M+H]⁺.

4.2.5.4. 2-(Chloromethyl)-1,4-bis(3,5-dichlorobenzyl)piperazine (**7d**). Prepared from (1,4-bis(3,5-dichlorobenzyl)piperazin-2-yl)methanol (**6d**) (0.50 g, 1.15 mmol). The product was obtained as a brown oil, yield 0.31 g (60%). TLC (95:5 CH₂Cl₂/MeOH), R_f = 0.99. ¹H NMR (DMSO-d₆): δ 7.33 (m, 6H, Ar), 3.94 (m, 2H, H-9_{a,b}), 3.85 (m, 2H,

H-7_{a,b}), 3.52 (m, 2H, H-8_{a,b}), 3.45 (m, 1H, H-2), 2.73 (m, 1H, H-3_a), 2.63 (m, 1H, H-3_b), 2.40 (m, 1H, H-6_a), 2.29 (m, 3H, H-6_b, H-5_{a,b}). ¹³C NMR (DMSO-d₆): δ 142.1 (2 x C, Ar), 141.3 (2 x C, Ar), 133.4 (2 x C, Ar), 130.5 (2 x CH, Ar), 128.7 (CH, Ar), 128.6 (CH, Ar), 127.7 (CH, Ar), 127.3 (CH, Ar), 61.5 (CH₂), 59.7 (CH, piperazine), 57.6 (CH₂), 56.7 (CH₂, piperazine), 54.9 (CH₂, piperazine), 52.8 (CH₂, piperazine), 43.4 (CH₂). [ESI-HRMS] calculated for C₁₉H₂₀Cl₅N₂: 451.0069 [M+H]⁺. Found: 451.0072 [M+H]⁺.

4.2.5.5. 2-(Chloromethyl)-1,4-bis(4-ethylbenzyl)piperazine (**7f**). Prepared from (1,4-bis(4-ethylbenzyl)piperazin-2-yl)methanol (**6f**) (1.35 g, 3.82 mmol). The product was purified by gradient column chromatography, eluting with petroleum ether –EtOAc 4:1 v/v to give the product as a brown solid, yield 1.05 g (74%). M.p. 80-82 °C. TLC (3:1 petroleum ether/EtOAc), R_f = 0.61. ¹H NMR (DMSO-d₆): δ 7.18 (m, 8H, Ar), 3.95 (m, 2H, H-9_{a,b}), 3.85 (m, 2H, H-7_{a,b}), 3.42 (m, 2H, H-8_{a,b}), 2.67 (m, 1H, H-2), 2.61 (m, 1H, H-3_a), 2.57 (q, *J* = 7.1 Hz, 4H, 2 x CH₂CH₃), 2.44 (m, 1H, H-3_b), 2.36 (m, 1H, H-6_a), 2.25 (m, 3H, H-6_b, H-5_{a,b}), 1.16 (t, *J* = 7.7 Hz, 6H, 2 x CH₃). ¹³C NMR (125 MHz, DMSO-d₆): δ 142.7 (2 x C, Ar), 136.2 (2 x C, Ar), 129.1 (4 x CH, Ar), 128.0 (4 x CH, Ar), 62.2 (CH₂), 59.7 (CH, piperazine), 57.2 (CH₂), 57.1 (CH₂, piperazine), 52.8 (CH₂, piperazine), 49.3 (CH₂, piperazine), 43.5 (CH₂), 28.29 (2 x CH₂CH₃), 16.08 (2 x CH₂CH₃). [ESI-HRMS] calculated for C₂₃H₃₂ClN₂: 371.2249 [M+H]⁺. Found: 371.2261 [M+H]⁺.

4.2.5.6. 2-(Chloromethyl)-1,4-bis(4-isopropylbenzyl)piperazine (**7g**). Prepared from (1,4-bis(4-isopropylbenzyl)piperazin-2-yl)methanol (**6g**) (1.50 g, 3.94 mmol). The product was purified by gradient column chromatography, eluting with petroleum ether –EtOAc 4:1 v/v to give the product as a brown solid, yield 1.37 g (88%). M.p. 64-66 °C. TLC (3:1 petroleum ether/EtOAc), R_f = 0.64. ¹H NMR (DMSO-d₆): δ 7.19 (m, 8H,

Ar), 3.95 (m, 2H, H-9_{a,b}), 3.85 (m, 2H, H-7_{a,b}), 3.41 (m, 2H, H-8_{a,b}), 2.85 (m, 2H, 2 x $\underline{\text{CH}}(\text{CH}_3)_2$), 2.68 (m, 1H, H-2), 2.60 (m, 1H, H-3_a), 2.44 (m, 1H, H-3_b), 2.36 (m, 1H, H-6_a), 2.24 (m, 3H, H-6_b, H-5_{a,b}), 1.19 (s, 6H, 2 x CH₃), 1.18 (s, 6H, 2 x CH₃). ¹³C NMR (DMSO-d₆): δ 147.5 (2 x C, Ar), 136.3 (2 x C, Ar), 129.1 (4 x CH, Ar), 126.5 (4 x CH, Ar), 62.2 (CH₂), 59.7 (CH, piperazine), 57.19 (CH₂), 55.4 (CH₂, piperazine), 52.8 (CH₂, piperazine), 49.2 (CH₂, piperazine), 43.4 (CH₂), 33.6 (2 x $\underline{\text{CH}}(\text{CH}_3)_2$), 24.4 (4 x CH₃). [ESI-HRMS] calculated for C₂₅H₃₆ClN₂: 399.2562 [M+H]⁺. Found: 399.2569 [M+H]⁺.

4.2.5.7. 2-(Chloromethyl)-1,4-bis(4-(tert-butyl)benzyl)piperazine (**7h**). Prepared from (1,4-bis(4-*tert*butylbenzyl)piperazin-2-yl)methanol (**6h**) (2.0 g, 4.89 mmol). The product was purified by gradient column chromatography, eluting with petroleum ether –EtOAc 4:1 v/v to give the product as a brown solid, yield 1.72 g (83%). M.p. 110-112 °C. TLC (3:1 petroleum ether/EtOAc), R_f = 0.66. ¹H NMR (DMSO-d₆): δ 7.25 (m, 8H, Ar), 3.97 (m, 2H, H-9_{a,b}), 3.43 (d, *J* = 13.2 Hz, 1H, H-7_a), 3.31 (d, *J* = 13.1 Hz, 1H, H-7_b), 2.79 (d, *J* = 12.6 Hz, 1H, H-8_a), 2.61 (d, *J* = 12.7 Hz, 1H, H-8_b), 2.46 (m, 3H, H-2, H-3_{a,b}), 2.22 (m, 4H, H-6_{a,b}, H-5_{a,b}), 1.28 (s, 9H, 3 x CH₃), 1.27 (s, 9H, 3 x CH₃). ¹³C NMR (DMSO-d₆): δ 149.3 (2 x C, Ar), 136.0 (2 x C, Ar), 128.9 (4 x CH, Ar), 125.1 (4 x CH, Ar), 61.9 (CH₂), 61.7 (CH, piperazine), 57.8 (CH₂), 56.6 (CH₂, piperazine), 53.2 (CH₂, piperazine), 50.7 (CH₂, piperazine), 43.5 (CH₂), 34.5 (2 x $\underline{\text{C}}(\text{CH}_3)_3$), 31.5 (6 x CH₃). [ESI-HRMS] calculated for C₂₇H₄₀ClN₂: 427.2875 [M+H]⁺. Found: 427.2889 [M+H]⁺.

4.2.5.8. 2-(Chloromethyl)-1,4-bis(pyridine-4-ylmethyl)piperazine (**7i**). Prepared from (1,4-bis(pyridine-4-yl)piperazin-2-yl)methanol (**6i**) (0.60 g, 2.01 mmol). The product was purified by gradient column chromatography eluting with petroleum ether –EtOAc 4:1 v/v to give the product as a colourless oil, yield 0.42 g (66%). TLC (2:1 petroleum ether/EtOAc), R_f = 0.59. ¹H NMR (DMSO-d₆): δ 8.50 (m, 4H, pyridine), 7.34 (m, 4H,

pyridine), 3.98 (m, 2H, H-9_{a,b}), 3.85 (d, $J = 13.9$ Hz, 1H, H-7_a), 3.55 (d, $J = 13.5$ Hz, 1H, H-7_b), 3.50 (m, 2H, H-8_{a,b}), 2.77 (m, 1H, H-2), 2.65 (m, 1H, H-3_a), 2.56 (m, 1H, H-3_b), 2.43 (m, 1H, H-6_a), 2.30 (m, 3H, H-6_b, H-5_{a,b}). ^{13}C NMR (DMSO- d_6): δ 150.0 (2 x CH, pyridine), 149.9 (2 x CH, pyridine), 148.6 (2 x C, pyridine), 124.2 (2 x CH, pyridine), 124.0 (2 x CH, pyridine), 60.9 (CH₂), 59.7 (CH, piperazine), 56.3 (CH₂), 55.4 (CH₂, piperazine), 52.8 (CH₂, piperazine), 49.2 (CH₂, piperazine), 43.3 (CH₂). [ESI-HRMS] calculated for C₁₇H₂₂ClN₄: 317.1533 [M+H]⁺. Found: 317.1546 [M+H]⁺.

4.2.5.9. 2-(Chloromethyl)-1,4-bis(pyridine-3-ylmethyl)piperazine (7j). Prepared from (1,4-bis(pyridine-3-yl)piperazin-2-yl)methanol (**6j**) (0.80 g, 2.68 mmol). The product was purified by gradient column chromatography eluting with petroleum ether–EtOAc 4:1 v/v to give the product as a colourless oil, yield 0.68 g (81%). TLC (2:1 petroleum ether/EtOAc), $R_f = 0.58$. ^1H NMR (DMSO- d_6): δ 8.48 (m, 4H, pyridine), 7.71 (t, $J = 5.4$ Hz, 2H, pyridine), 7.35 (d, $J = 5.5$ Hz, 2H, pyridine), 3.98 (m, 2H, H-9_{a,b}), 3.87 (d, $J = 13.4$ Hz, 1H, H-7_a), 3.54 (d, $J = 13.5$ Hz, 1H, H-7_b), 3.47 (m, 2H, H-8_{a,b}), 2.72 (m, 1H, H-2), 2.62 (m, 1H, H-3_a), 2.40 (m, 1H, H-3_b), 2.30 (m, 2H, H-6_{a,b}), 2.13 (m, 2H, H-5_{a,b}). ^{13}C NMR (DMSO- d_6): δ 150.4 (CH, pyridine), 150.3 (CH, pyridine), 148.8 (CH, pyridine), 148.7 (CH, pyridine), 136.9 (CH, pyridine), 136.8 (CH, pyridine), 134.6 (2 x C, pyridine), 123.9 (2 x CH, pyridine), 62.2 (CH₂), 60.3 (CH, piperazine), 55.5 (CH₂), 54.8 (CH₂, piperazine), 51.3 (CH₂, piperazine), 48.9 (CH₂, piperazine), 43.2 (CH₂). [ESI-HRMS] calculated for C₁₇H₂₂ClN₄: 317.1533 [M+H]⁺. Found: 317.1540 [M+H]⁺.

4.2.6. General method for the preparation of imidazole derivatives (8). To a stirred suspension of K₂CO₃ (4 m.eq.) in dry acetonitrile (10 mL/4 mmol of K₂CO₃) was added imidazole (4 m.eq.). The reaction mixture was refluxed at 45 °C for 1 h. After cooling to room temperature the chloride (**7**) (1 m.eq.) was added and the reaction mixture refluxed at 70 °C for 48 h. The solvent was evaporated under reduced pressure and the

resulting mixture diluted with EtOAc (50 mL/mmol of chloride (**7**) used) and washed with brine (3 x 20 mL/mmol) and H₂O (3 x 20 mL/mmol). The organic layer was dried (MgSO₄) and evaporated under reduced pressure to give the crude imidazole, which was purified by gradient column chromatography.

4.2.6.1. *2-((1H-Imidazol-1-yl)methyl)-1,4-bis(4-fluorobenzyl)piperazine* (**8a**).

Prepared from 2-(chloromethyl)-1,4-bis(4-fluorobenzyl)piperazine (**7a**) (0.50 g, 1.42 mmol). The product was purified by gradient column chromatography eluting with CH₂Cl₂-MeOH 98:2 v/v, followed by further purification by preparative TLC (CH₂Cl₂-MeOH 95:5 to 9:1 v/v) to give the product as a yellow oil*, yield 0.29 g (53%). TLC (95:5 CH₂Cl₂/MeOH), R_f = 0.50. ¹H NMR (DMSO-d₆): δ 7.47 (s, 1H, imidazole), 7.36-7.11 (m, 8H, Ar), 6.97 (s, 1H, imidazole), 6.82 (s, 1H, imidazole), 4.30 (dd, *J* = 4.1, 13.6 Hz, 1H, H-9_a), 4.22 (dd, *J* = 7.0, 13.6 Hz, 1H, H-9_b), 3.93 (d, *J* = 13.6 Hz, 1H, H-7_a), 3.59 (d, *J* = 13.2 Hz, 1H, H-7_b), 3.44 (d, *J* = 13.2 Hz, 1H, H-8_a), 3.32 (d, *J* = 13.2 Hz, 1H, H-8_b), 2.82 (m, 1H, H-2), 2.74 (m, 1H, H-3_a), 2.30 (m, 4H, H-3_b, H-6_{a,b}, H-5_a), 2.12 (m, 1H, H-5_b). ¹³C NMR (DMSO-d₆): δ 139.3 (2 x C, Ar), 134.7 (2 x C, Ar), 131.2 (2 x CH, Ar), 131.1 (2 x CH, Ar), 130.7 (CH, imidazole), 128.9 (2 x CH, Ar), 128.7 (2 x CH, Ar), 127.3 (CH, imidazole), 120.3 (CH, imidazole), 61.6 (CH₂), 59.3 (CH, piperazine), 57.7 (CH₂), 56.8 (CH₂), 55.4 (CH₂, piperazine), 52.3 (CH₂, piperazine), 44.3 (CH₂, piperazine). ¹⁹F NMR (DMSO-d₆): δ -115.87 (2 x C₆H₄F). [ESI-HRMS] calculated for C₂₂H₂₅F₂N₄: 383.2047 [M+H]⁺. Found: 383.2050 [M+H]⁺.

* The product was found to be unstable (light/moisture) so was wrapped in aluminium foil and stored in the freezer.

4.2.6.2. *2-((1H-Imidazol-1-yl)methyl)-1,4-bis(4-chlorobenzyl)piperazine* (**8b**).

Prepared from 2-(chloromethyl)-1,4-bis(4-chlorobenzyl)piperazine (**7b**) (0.15 g, 0.39 mmol). The product was purified by gradient column chromatography eluting with

CH₂Cl₂-MeOH 97.5:2.5 v/v, followed by further purification by preparative TLC (CH₂Cl₂-MeOH 95:5 v/v) to give the product as a colourless oil, yield 0.09 g (56%). TLC (95:5 CH₂Cl₂/MeOH), *R_f* = 0.60. ¹H NMR (CDCl₃): δ 7.24 (s, 1H, imidazole), 7.22-7.12 (m, 8H, Ar), 6.89 (s, 1H, imidazole), 6.59 (s, 1H, imidazole), 4.28 (dd, *J* = 4.3, 13.6 Hz, 1H, H-9_a), 4.22 (dd, *J* = 7.7, 13.6 Hz, 1H, H-9_b), 3.78 (d, *J* = 13.0 Hz, 1H, H-7_a), 3.54 (d, *J* = 13.0 Hz, 1H, H-7_b), 3.36 (d, *J* = 13.1 Hz, 1H, H-8_a), 3.27 (d, *J* = 12.9 Hz, 1H, H-8_b), 2.80 (m, 1H, H-2), 2.76 (m, 1H, H-3_a), 2.42 (m, 1H, H-3_b), 2.40-2.13 (m, 4H, H-6_{a,b}, H-5_{a,b}). ¹³C NMR (CDCl₃): δ 138.1 (C, Ar), 136.9 (C, Ar), 136.4 (C, Ar), 133.0 (C, Ar), 130.4 (CH, imidazole), 128.6 (4 x CH, Ar), 128.5 (4 x CH, Ar), 127.3 (CH, imidazole), 120.5 (CH, imidazole), 62.1 (CH₂), 58.7 (CH, piperazine), 58.3 (CH₂), 53.8 (CH₂), 52.2 (CH₂, piperazine), 48.3 (CH₂, piperazine), 44.1 (CH₂, piperazine). [ESI-HRMS] calculated for C₂₂H₂₅Cl₂N₄: 415.1456 [M+H]⁺. Found: 415.1447 [M+H]⁺.

4.2.6.3. 2-((1*H*-Imidazol-1-yl)methyl)-1,4-bis(4-bromobenzyl)piperazine (**8c**).

Prepared from 2-(chloromethyl)-1,4-bis(4-bromobenzyl)piperazine (**7c**) (0.4 g, 0.84 mmol). The product was purified by gradient column chromatography eluting with CH₂Cl₂-MeOH 95:5 v/v to give the product as a white solid, yield 0.26 g (62%). M.p. 54-56 °C. TLC (95:5 CH₂Cl₂/MeOH), *R_f* = 0.68. ¹H NMR (CDCl₃): δ 7.51 (s, 1H, imidazole), 7.49 (m, 4H, Ar), 7.26 (d, *J* = 8.2 Hz, 2H, Ar), 7.21 (d, *J* = 8.2 Hz, 2H, Ar), 7.00 (s, 1H, imidazole), 6.83 (s, 1H, imidazole), 4.31 (dd, *J* = 4.6, 13.5 Hz, 1H, H-9_a), 4.20 (dd, *J* = 7.0, 13.5 Hz, 1H, H-9_b), 3.89 (d, *J* = 14.0 Hz, 1H, H-7_a), 3.57 (d, *J* = 14.2 Hz, 1H, H-7_b), 3.43 (d, *J* = 13.4 Hz, 1H, H-8_a), 3.32 (d, *J* = 13.7 Hz, 1H, H-8_b), 2.82 (m, 1H, H-2), 2.75 (m, 1H, H-3_a), 2.30 (m, 4H, H-3_b, H-6_{a,b}, H-5_a), 2.10 (m, 1H, H-5_b). ¹³C NMR (CDCl₃): δ 139.0 (2 x C, Ar), 138.0 (2 x C, Ar), 131.5 (CH, imidazole), 131.4 (4 x CH, Ar), 131.0 (4 x CH, Ar), 128.6 (CH, imidazole), 120.4 (CH, imidazole), 61.7

(CH₂), 59.3 (CH, piperazine), 56.8 (CH₂), 54.1 (CH₂), 52.2 (CH₂, piperazine), 46.5 (CH₂, piperazine), 44.4 (CH₂, piperazine). Anal. Calcd for C₂₂H₂₄Br₂N₄ (504.2608): C, 52.40; H, 4.80; N, 11.11. Found: C, 52.30; H, 4.91; N, 11.20.

4.2.6.4. 2-((1H-Imidazol-1-yl)methyl)-1,4-bis(3,5-dichlorobenzyl)piperazine (8d).

Prepared from 2-(chloromethyl)-1,4-bis(3,5-dichlorobenzyl)piperazine (**7d**) (0.35 g, 0.77 mmol). The product was purified by gradient column chromatography eluting with CH₂Cl₂-MeOH 99.8:0.2 v/v followed by further purification by preparative TLC (petroleum ether-EtOAc 3:2 v/v) to give the product as a colourless oil, yield 0.18 g (51%). TLC (1:1 petroleum ether/EtOAc), R_f = 0.94. ¹H NMR (CDCl₃): δ 7.47 (s, 1H, imidazole), 7.28 (s, 1H, para-Ar), 7.26 (s, 1H, para-Ar), 7.21 (s, 2H, Ar), 7.19 (s, 2H, Ar), 7.05 (s, 1H, imidazole), 6.77 (s, 1H, imidazole), 4.29 (dd, *J* = 4.1, 13.6 Hz, 1H, H-9_a), 4.21 (dd, *J* = 7.0, 13.6 Hz, 1H, H-9_b), 3.79 (d, *J* = 14.0 Hz, 1H, H-7_a), 3.64 (d, *J* = 13.9 Hz, 1H, H-7_b), 3.46 (d, *J* = 13.4 Hz, 1H, H-8_a), 3.39 (d, *J* = 13.4 Hz, 1H, H-8_b), 2.96 (m, 1H, H-2), 2.88 (m, 1H, H-3_a), 2.52 (m, 3H, H-3_b, H-6_{a,b}), 2.35 (m, 2H, H-5_{a,b}). ¹³C NMR (CDCl₃): δ 142.1 (2 x C, Ar), 141.5 (2 x C, Ar), 135.1 (2 x C, Ar), 129.3 (CH, imidazole), 127.6 (2 x CH, Ar), 127.2 (2 x CH, Ar), 126.9 (CH, imidazole), 126.6 (2 x CH, Ar), 119.4 (CH, imidazole), 61.8 (CH₂), 59.0 (CH, piperazine), 57.4 (CH₂), 53.7 (CH₂), 52.0 (CH₂, piperazine), 47.8 (CH₂, piperazine), 44.2 (CH₂, piperazine). [ESI-HRMS] calculated for C₂₂H₂₃Cl₄N₄: 483.0677 [M+H]⁺. Found: 483.0678 [M+H]⁺.

4.2.6.5. 2-((1H-Imidazol-1-yl)methyl)-1,4-bis(4-methoxybenzyl)piperazine (8e).

Prepared from 2-(chloromethyl)-1,4-bis(4-methoxybenzyl)piperazine (**7e**) (0.35 g, 0.93 mmol). The product was purified by gradient column chromatography eluting with CH₂Cl₂-MeOH 97.5:2.5 v/v followed by further purification by preparative TLC (CH₂Cl₂-MeOH 9:1 v/v) to give the product as a yellow oil, yield 0.25 g (68%). TLC (95:5 CH₂Cl₂/MeOH), R_f = 0.53. ¹H NMR (DMSO-d₆): δ 7.46 (s, 1H, imidazole), 7.19

(d, $J = 8.5$ Hz, 2H, Ar), 7.18 (d, $J = 8.5$ Hz, 2H, Ar), 6.96 (s, 1H, imidazole), 6.87 (d, $J = 8.2$ Hz, 4H, Ar), 6.81 (s, 1H, imidazole), 4.29 (dd, $J = 4.4, 13.9$ Hz, 1H, H-9_a), 4.20 (dd, $J = 7.8, 13.9$ Hz, 1H, H-9_b), 3.84 (d, $J = 13.0$ Hz, 1H, H-7_a), 3.73 (s, 6H, 2 x OCH₃), 3.51 (d, $J = 12.9$ Hz, 1H, H-7_b), 3.39 (d, $J = 12.8$ Hz, 1H, H-8_a), 3.25 (d, $J = 12.8$ Hz, 1H, H-8_b), 2.77 (m, 1H, H-2), 2.69 (m, 1H, H-3_a), 2.30-2.22 (m, 4H, H-3_b, H-6_{a,b}, H-5_a), 2.09 (m, 1H, H-5_b). ¹³C NMR (DMSO-d₆): δ 142.8 (C, Ar), 142.7 (C, Ar), 134.6 (C, Ar), 131.0 (C, Ar), 130.5 (2 x CH, Ar), 130.1 (2 x CH, Ar), 128.6 (CH, imidazole), 128.1 (CH, imidazole), 120.1 (CH, imidazole), 114.1 (2 x CH, Ar), 114.0 (2 x CH, Ar), 62.0 (CH₂), 59.2 (CH, piperazine), 58.0 (CH₂), 57.0 (CH₂), 55.5 (2 x OCH₃), 54.8 (CH₂, piperazine), 54.3 (CH₂, piperazine), 52.3 (CH₂, piperazine). [ESI-HRMS] calculated for C₂₄H₃₁N₄O₂: 407.2442 [M+H]⁺. Found: 407.2438 [M+H]⁺.

4.2.6.6. 2-((1H-imidazol-1-yl)methyl)-1,4-bis(4-ethylbenzyl)piperazine (8f). Prepared from 2-(chloromethyl)-1,4-bis(4-ethylbenzyl)piperazine (**7f**) (1.50 g, 4.04 mmol). The product was purified by gradient column chromatography eluting with CH₂Cl₂-MeOH 99.5:0.5 v/v followed by further purification by preparative TLC (CH₂Cl₂-MeOH 95:5 v/v) to give the product as a yellow oil, yield 1.12 g (69%). TLC (95:5 CH₂Cl₂/MeOH), $R_f = 0.51$. ¹H NMR (DMSO-d₆): δ 7.45 (s, 1H, imidazole), 7.16 (m, 8H, Ar), 6.93 (s, 1H, imidazole), 6.80 (s, 1H, imidazole), 4.29 (dd, $J = 4.3, 13.8$ Hz, 1H, H-9_a), 4.21 (dd, $J = 7.7, 13.8$ Hz, 1H, H-9_b), 3.88 (d, $J = 13.3$ Hz, 1H, H-7_a), 3.54 (d, $J = 13.3$ Hz, 1H, H-7_b), 3.4 (d, $J = 13.1$ Hz, 1H, H-8_a), 3.27 (d, $J = 12.9$ Hz, 1H, H-8_b), 2.78 (m, 1H, H-2), 2.71 (m, 1H, H-3_a), 2.58 (m, 4H, 2 x CH₂CH₃), 2.31-2.24 (m, 3H, H-3_b, H-6_{a,b}), 2.11 (m, 2H, H-5_{a,b}), 1.17 (t, $J = 7.58$ Hz, 3H, CH₂CH₃), 1.16 (t, $J = 7.6$ Hz, 3H, CH₂CH₃). ¹³C NMR (DMSO-d₆): δ 142.9 (C, Ar), 142.7 (C, Ar), 136.5 (C, Ar), 135.7 (C, Ar), 129.4 (2 x CH, Ar), 128.9 (2 x CH, Ar), 128.5 (CH, imidazole), 128.1 (2 x CH, Ar), 128.0 (2 x CH, Ar), 126.4 (CH, imidazole), 120.3 (CH, imidazole), 62.3 (CH₂), 59.3

(CH, piperazine), 58.8 (CH₂), 57.4 (CH₂), 55.4 (CH₂, piperazine), 54.3 (CH₂, piperazine), 52.4 (CH₂, piperazine), 28.3 (CH₂CH₃), 28.2 (CH₂CH₃), 16.2 (CH₂CH₃), 16.1 (CH₂CH₃). [ESI-HRMS] calculated for C₂₆H₃₅N₄: 403.2862 [M+H]⁺. Found: 403.2857 [M+H]⁺.

4.2.6.7. 2-((1*H*-Imidazol-1-yl)methyl)-1,4-bis(4-isopropylbenzyl)piperazine (**8g**).

Prepared from 2-(chloromethyl)-1,4-bis(4-isopropylbenzyl)piperazine (**7g**) (1.50 g, 3.75 mmol). The product was purified by gradient column chromatography eluting with CH₂Cl₂-MeOH 99.7:0.3 v/v followed by further purification by preparative TLC (petroleum ether-EtOAc 3:2 v/v) to give the product as a brown solid, yield 1.02 g (63%). M.p. 110-112 °C. TLC (2:1 petroleum ether/EtOAc), R_f = 0.53. ¹H NMR (DMSO-d₆): δ 7.43 (s, 1H, imidazole), 7.19 (m, 8H, Ar), 6.90 (s, 1H, imidazole), 6.79 (s, 1H, imidazole), 4.28 (dd, *J* = 4.4, 14.0 Hz, 1H, H-9_a), 4.21 (dd, *J* = 7.8, 14.0 Hz, 1H, H-9_b), 3.87 (d, *J* = 13.3 Hz, 1H, H-7_a), 3.55 (d, *J* = 13.3 Hz, 1H, H-7_b), 3.43 (d, *J* = 13.0 Hz, 1H, H-8_a), 3.26 (d, *J* = 13.0 Hz, 1H, H-8_b), 2.85 (m, 2H, 2 x CH(CH₃)₂), 2.78 (m, 1H, H-2), 2.71 (m, 1H, H-3_a), 2.32-2.24 (m, 3H, H-3_b, H-6_{a,b}), 2.10 (m, 2H, H-5_{a,b}), 1.98 (d, *J* = 6.9 Hz, 6H, 2 x CH₃), 1.89 (d, *J* = 6.8 Hz, 6H, 2 x CH₃). ¹³C NMR (DMSO-d₆): δ 147.5 (C, Ar), 147.4 (C, Ar), 136.6 (C, Ar), 135.9 (C, Ar), 129.4 (2 x CH, Ar), 128.9 (2 x CH, Ar), 128.5 (CH, imidazole), 126.6 (2 x CH, Ar), 126.5 (2 x CH, Ar), 126.4 (CH, imidazole), 120.3 (CH, imidazole), 62.3 (CH₂), 59.4 (CH, piperazine), 57.4 (CH₂), 56.9 (CH₂), 55.4 (CH₂, piperazine), 54.3 (CH₂, piperazine), 52.5 (CH₂, piperazine), 33.6 (2 x CH(CH₃)₂), 24.4 (4 x CH₃). [ESI-HRMS] calculated for C₂₈H₃₉N₄: 431.3175 [M+H]⁺. Found: 431.3186 [M+H]⁺.

4.2.6.8. 2-((1*H*-Imidazol-1-yl)methyl)-1,4-bis(4-(*tert*-butyl)benzyl)piperazine (**8h**).

Prepared from 2-(chloromethyl)-1,4-bis(4-*tert*butylbenzyl)piperazine (**7h**) (1.50 g, 3.51 mmol). The product was purified by gradient column chromatography eluting with

CH₂Cl₂-MeOH 99.8:0.2 v/v followed by further purification by preparative TLC (petroleum ether-EtOAc 3:2 v/v) to give the product as a yellow oil, yield 0.85 g (53%). TLC (2:1 petroleum ether/EtOAc), *R_f* = 0.55. ¹H NMR (DMSO-*d*₆): δ 7.43 (s, 1H, imidazole), 7.33 (m, 4H, Ar), 7.20 (m, 4H, Ar), 6.89 (s, 1H, imidazole), 6.78 (s, 1H, imidazole), 4.28 (dd, *J* = 4.3, 13.8 Hz, 1H, H-9_a), 4.21 (dd, *J* = 7.6, 13.8 Hz, 1H, H-9_b), 3.87 (d, *J* = 13.7 Hz, 1H, H-7_a), 3.56 (d, *J* = 13.7 Hz, 1H, H-7_b), 3.44 (d, *J* = 12.8 Hz, 1H, H-8_a), 3.27 (d, *J* = 12.8 Hz, 1H, H-8_b), 2.80 (m, 1H, H-2), 2.72 (m, 1H, H-3_a), 2.36 (m, 1H, H-3_b), 2.29 (m, 2H, H-6_{a,b}), 2.11 (m, 2H, H-5_{a,b}), 1.28 (s, 9H, 3 x CH₃), 1.27 (s, 9H, 3 x CH₃). ¹³C NMR (DMSO-*d*₆): δ 149.8 (C, Ar), 149.6 (C, Ar), 136.2 (C, Ar), 135.5 (C, Ar), 129.1 (2 x CH, Ar), 128.6 (2 x CH, Ar), 128.5 (CH, imidazole), 125.4 (2 x CH, Ar), 125.3 (2 x CH, Ar), 125.2 (CH, imidazole), 120.3 (CH, imidazole), 62.2 (CH₂), 59.4 (CH, piperazine), 57.4 (CH₂), 56.3 (CH₂), 55.4 (CH₂, piperazine), 54.3 (CH₂, piperazine), 52.5 (CH₂, piperazine), 34.6 (2 x C(CH₃)₃), 31.7 (6 x CH₃). [ESI-HRMS] calculated for C₃₀H₄₃N₄: 459.3488 [M+H]⁺. Found: 459.3490 [M+H]⁺.

4.2.6.9. 2-((1*H*-Imidazol-1-yl)methyl)-1,4-bis(pyridine-4-ylmethyl)piperazine (8i).
 Prepared from 2-(chloromethyl)-1,4-bis(pyridine-4-ylmethyl)piperazine (**7i**) (0.4 g, 1.26 mmol). The product was purified by gradient column chromatography, eluting with CH₂Cl₂-MeOH-Et₃N 94:5:1 v/v/v, followed by further purification by preparative TLC (CH₂Cl₂-MeOH 9:1 to 85:15 v/v) to give the product as a yellow oil, yield 0.26 g (58%). TLC (1:9 CH₂Cl₂/MeOH), *R_f* = 0.57. ¹H NMR (DMSO-*d*₆): δ 8.47 (m, 4H, pyridine), 7.33 (s, 1H, imidazole), 7.15 (m, 4H, pyridine), 6.94 (s, 1H, imidazole), 6.68 (s, 1H, imidazole), 4.28 (dd, *J* = 4.4, 14.0 Hz, 1H, H-9_a), 4.21 (dd, *J* = 7.8, 14.0 Hz, 1H, H-9_b), 3.79 (d, *J* = 14.0 Hz, 1H, H-7_a), 3.58 (d, *J* = 14.0 Hz, 1H, H-7_b), 3.49 (d, *J* = 13.4 Hz, 1H, H-8_a), 3.40 (d, *J* = 13.4 Hz, 1H, H-8_b), 2.88 (m, 1H, H-2), 2.83 (m, 1H, H-3_a), 2.45-2.21 (m, 5H, H-3_b, H-6_{a,b}, H-5_{a,b}). ¹³C NMR (CDCl₃): δ 150.0 (2 x CH, pyridine),

149.9 (2 x CH, pyridine), 137.5 (CH, imidazole), 137.3 (C, pyridine), 137.0 (C, pyridine), 129.5 (CH, imidazole), 123.8 (2 x CH, pyridine), 123.2 (2 x CH, pyridine), 119.3 (CH, imidazole), 61.7 (CH₂), 59.2 (CH, piperazine), 57.3 (CH₂), 54.0 (CH₂), 52.0 (CH₂, piperazine), 48.1 (CH₂, piperazine), 44.5 (CH₂, piperazine). [ESI-HRMS] calculated for C₂₀H₂₅N₆: 349.2141 [M+H]⁺. Found: 349.2146 [M+H]⁺.

4.2.6.10. 2-((1*H*-Imidazol-1-yl)methyl)-1,4-bis(pyridine-3-ylmethyl)piperazine (8j).

Prepared from 2-(chloromethyl)-1,4-bis(pyridine-3-ylmethyl)piperazine (**7j**) (0.44 g, 1.41 mmol). The product was purified by gradient column chromatography, eluting with CH₂Cl₂-MeOH-Et₃N 94:5:1 v/v/v, followed by further purification by preparative TLC (CH₂Cl₂-MeOH 9:1 to 85:15 v/v) to give the product as a yellow oil, yield 0.25 g (53%). TLC (1:9 CH₂Cl₂/MeOH), R_f = 0.56. ¹H NMR (DMSO-d₆): δ 8.50 (s, 1H, pyridine), 8.49 (s, 1H, pyridine), 8.46 (m, 2H, pyridine), 7.71 (m, 2H, pyridine), 7.61 (s, 1H, imidazole), 7.50 (s, 1H, imidazole), 7.34 (m, 2H, pyridine), 6.82 (s, 1H, imidazole), 4.29 (dd, *J* = 4.4, 13.8 Hz, 1H, H-9_a), 4.21 (dd, *J* = 7.7, 13.8 Hz, 1H, H-9_b), 3.94 (d, *J* = 13.8 Hz, 1H, H-7_a), 3.64 (d, *J* = 13.8 Hz, 1H, H-7_b), 3.49 (d, *J* = 13.4 Hz, 1H, H-8_a), 3.39 (d, *J* = 13.4 Hz, 1H, H-8_b), 2.86 (m, 1H, H-2), 2.75 (m, 2H, H-3_{a,b}), 2.32 (m, 3H, H-6_{a,b}, H-5_a), 2.13 (m, 1H, H-5_b). ¹³C NMR (DMSO-d₆): δ 150.2 (CH, pyridine), 149.6 (CH, pyridine), 148.6 (CH, pyridine), 148.5 (CH, pyridine), 137.3 (C, pyridine), 137.0 (C, pyridine), 136.4 (CH, imidazole), 135.3 (CH, pyridine), 129.0 (CH, pyridine), 124.0 (CH, imidazole), 121.8 (2 x CH, pyridine), 119.4 (CH, imidazole), 60.1 (CH₂), 58.8 (CH, piperazine), 55.5 (CH₂), 53.3 (CH₂), 51.9 (CH₂, piperazine), 47.6 (CH₂, piperazine), 44.1 (CH₂, piperazine). [ESI-HRMS] calculated for C₂₀H₂₅N₆: 349.2141 [M+H]⁺. Found: 349.2149 [M+H]⁺.

4.3. CYP121A1 Spectral binding assays for ligand K_D determination

CYP121A1 protein was expressed and purified as described previously.²⁵ Ligand binding assays were performed by spectrophotometric titration using a Cary 60 UV-visible scanning spectrophotometer (Agilent, UK) and a 1 cm path length quartz cuvette, recording spectra between 250 and 800 nm. Titrations were typically done with 3-5 μ M CYP121A1 at 25 °C in 100 mM potassium phosphate (KPi) buffer, 200 mM KCl, pH 7.85 with 0.004% Triton X-100. Ligand stock solutions were prepared in dimethylsulfoxide (DMSO). Ligands were added in small volumes (typically 0.05-0.2 μ L aliquots) from concentrated stock solutions to the protein in a 1 mL final volume. Spectral measurements were taken before ligand addition, and following addition of each aliquot of ligand until no further spectral change occurred. Difference spectra at each stage in the titration were obtained by subtraction of the initial ligand-free enzyme spectrum from subsequent spectra collected after each addition of ligand. From the difference spectra, a pair of wavelengths were identified and defined as the absorbance maximum (A_{peak}) and minimum (A_{trough}). The overall absorbance change (ΔA_{max}) was calculated by subtracting the A_{trough} value from the A_{peak} value for each spectrum collected after a ligand addition. Graphs of ΔA_{max} against [ligand] were plotted for each titration. Titrations were done in triplicate and the final K_D value presented was determined as the average value across the three sets. The K_D values were determined by fitting the data using either a standard hyperbolic function (Equation 1) or the Hill equation (Equation 2) using Origin software (OriginLab, Northampton, MA).

$$A_{\text{obs}} = (A_{\text{max}} * L / (K_d + L)) \quad (\text{Equation 1})$$

In Equation 1 (the standard hyperbolic function, the Michaelis-Menten function adapted for ligand binding), A_{obs} is the observed absorbance change at ligand concentration L , A_{max} is the maximal absorbance change observed at apparent ligand

saturation, and K_d is the dissociation constant for the binding of the ligand (the substrate concentration at which $A_{obs} = 0.5 \times A_{max}$).

$$A_{obs} = (A_{max} \times L^n)/(K^n + L^n) \quad (\text{Equation 2})$$

In Equation 2 (the sigmoidal Hill equation), A_{obs} is the observed absorbance change at ligand concentration L , A_{max} is the absorbance change at apparent ligand saturation, K is the apparent dissociation constant, and n is the Hill coefficient, a value describing the apparent extent of cooperativity observed in ligand binding.

Antimycobacterial activity assay: *M. tuberculosis* H₃₇Rv was grown in 7H9 liquid medium with 10 % Middlebrook OADC Growth Supplement enrichment (BBL/Becton-Dickinson, Sparks, MD, USA). Cells were cultured at 37 °C until mid-log phase was reached ($OD_{600nm} = 0.4-0.6$). After cells reached mid-log phase, bacterial suspensions were prepared as described below and REMA assays were performed. The anti-*M. tuberculosis* activities of the compounds were determined by the REMA (Resazurin Microtiter Assay) method.²⁶ Stock solutions of the tested compounds (10 mg/mL) were prepared in DMSO and diluted in Middlebrook 7H9 broth supplemented with 10% OADC. The microdilution of the compounds was performed in 96-well plates to obtain final compound concentration ranges of 0.39-100 µg/mL. Rifampicin in the concentration range between 0.004-1 µg/mL was added as control. Bacterial suspensions were prepared and their turbidity adjusted to match the optical density of McFarland no. 1 standard. After a further dilution of 1:20 in Middlebrook 7H9 broth supplemented with OADC, 100 µL of the inoculum were added to each well of the 96-well plate. Cultures were incubated for 7 days at 37 °C, and 30 µL of 0.01% resazurin was added. Wells were read after 24 h for colour change and measured as the fluorescence (excitation/emission of 530/590 nm filters, respectively) in a

microfluorimeter. The MIC was defined as the lowest concentration resulting in 90 % inhibition of *M. tuberculosis* growth. The presented results are representative from two independent experiments.

4.4. Computational methods

4.4.1 Molecular Modeling and Docking

Docking studies were performed using the MOE²² program and Mtb CYP121A1 co-crystallised with cYY (PDB 3G5H). All minimisations were performed with MOE until a RMSD gradient of 0.01 Kcal/mol/Å with the MMFF94 forcefield and partial charges were automatically calculated. The charge of the haem iron at physiological pH was set to 3⁺ (geometry d2sp3) through the atom manager in MOE. The Alpha Triangle placement, which derives poses by random superposition of ligand atom triplets through alpha sphere dummies in the receptor site, was chosen to determine the poses. The London ΔG scoring function estimates the free energy of binding of the ligand from a given pose. Refinement of the results was done using the MMFF94 forcefield, and rescoring of the refined results using the London ΔG scoring function was applied. The output database dock file was created with different poses for each ligand and arranged according to the final score function (S), which is the score of the last stage that was not set to zero.

4.4.2 Molecular Dynamics Simulation

Molecular dynamics simulations were run on the CYP121A1 protein in complex with **8g** and **8h**. Both PDB files were first optimised with protein preparation wizard in Maestro,²⁷ version 11.8.012 by assigning bond orders, adding hydrogen, and correcting incorrect bond types. A default quick relaxation protocol was used to minimise the MD systems with the Desmond programme.²⁸ In Desmond, the volume of space in which the simulation takes place, the global cell, is built up by regular 3D simulation boxes, which was utilised as part in this system for

protein interactions. The orthorhombic water box allowed for a 10 Å buffer region between protein atoms and box sides. Overlapping water molecules were deleted, and the systems were neutralised with Na⁺ ions and salt concentration 0.15 M. Force-field parameters for the complexes were assigned using the OPLS_2005 forcefield, that is, a 50ns molecular dynamic run in the NPT ensemble (T = 300 K) at a constant pressure of 1 bar. Energy and trajectory atomic coordinate data were recorded at each 1.2 ns.

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A. Supplementary data

Supplementary data associated with this article can be found, in the online version, at

References:

- (1) World Health Organization. Global tuberculosis report **2018**. Available from: http://www.who.int/tb/publications/global_report/en/ (accessed January 3rd, 2019).
- (2) Mclean, K.J.; Munro, A.W. Structural biology and biochemistry of cytochrome P450 systems in *Mycobacterium tuberculosis*. *Drug. Metab. Rev.* **2008**, *40*, 427-446.
- (3) Nunn, A.J.; Jindani, A.; Enarson, D.A. Results at 30 months of a randomised trial of two 8-month regimens for the treatment of tuberculosis. *Int. J. Tuberc. Lung Dis.* **2011**, *15*, 741-745.
- (4) Stagg, H.R.; Harris, R.J.; Hatherell, H.A.; Obach, D.; Zhao, H.; Tsuchiya, N.; Kranzer, K.; Nikolayevskyy, Y.; Kim, J.; Lipman, M.C.; Abubaker, I. What are the most efficacious treatment regimens for isoniazid-resistant tuberculosis? A systematic review and network meta-analysis. *Thorax*. **2016**, *71*, 940-949.
- (5) Falzon, D.; Schünemann, H.J.; Harausz, E.; González-Angulo, L.; Lienhardt, C.; Jaramillo, E.; Weyer, K. World Health Organization treatment guidelines for drug resistant tuberculosis, 2016 Update. *Eur. Respir. J.* **2017**, *49*, 1602308.
- (6) Shleeve, M.O.; Kudykina, Y.K.; Vostroknutova, G.N.; Suzina, N.E.; Mulyukin, A.L.; Kaprelyants, A.S. Dormant ovoid cells of *Mycobacterium tuberculosis* are formed in response to gradual external acidification. *Tuberculosis* **2011**, *91*, 146-154.
- (7) Zhang, Y.; Yew, W.W.; Barer, M.R. Targeting persisters for tuberculosis control. *Antimicrob. Agents Chemother.* **2012**, *56*, 2223-2230.

- (8) Floyd, K.; Glaziou, P.; Zumla, A.; Raviglione, M. The global tuberculosis epidemic and progress in care, prevention, and research: an overview in year 3 of the End TB era. *Lancet* **2018**, *6*, 299-314.
- (9) Cole, S.T.; Brosch, R.; Parkhill, J.; Garnier, T.; Churcher, C.; Harris, D.; Gordon, S. V.; Eiglmeier, K.; Gas, S.; Barry, C. E.; Tekaia, F.; Badcock, K.; Basham, D.; Brown, D.; Chillingworth, T.; Connor, R.; Davies, R.; Devlin, K.; Feltwell, T.; Gentles, S.; Hamlin, N.; Holroyd, S.; Hornby, T.; Jagels, K.; Krogh, A.; McLean, J.; Moule, S.; Murphy, L.; Oliver, K.; Osborne, J.; Quail, M. A.; Rajandream, M. A.; Rogers, J.; Rutter, S.; Seeger, K.; Skelton, J.; Squares, R.; Squares, S.; Sulston, J.; Taylor, K.; Whitehead, S.; Barrell, B. Deciphering the biology of *Mycobacterium tuberculosis* from the complete genome sequence. *Nature* **1998**, *393*, 537-544.
- (10) Mclean, K.J.; Clift, D.; Lewis, D.G.; Sabri, M.; Balding, P.R.; Sutcliffe, M.J.; Leys, D.; Munro, A.W. The preponderance of P450s in the *Mycobacterium tuberculosis* genome. *Trends Microbiol.* **2006**, *14*, 220-228.
- (11) (64) Megehee, J.A.; Lundrigan, M.D. Temporal expression of *Mycobacterium smegmatis* respiratory terminal oxidases. *Can. J. Microbiol.* **2007**, *53*, 459-463.
- (12) Sassetti C.M.; Boyd, D.H.; Rubin, E.J. Genes required for mycobacterial growth defined by high density mutagenesis. *Mol. Microbiol.* **2003**, *48*, 77-84.
- (13) Sassetti C.M.; Boyd, D.H.; Rubin, E.J. Comprehensive identification of conditionally essential genes in mycobacteria. *Proc. Natl. Acad. Sci.* **2001**, *98*, 12712-12717.
- (14) Mclean, K.J.; Carroll, P.; Lewis, D.G.; Dunford, A.; Seward, H.E.; Neeli, R.; Cheesman, M.R.; Marsollier, L.; Douglas, P.; Smith, W.E.; Rosenkrands, I.; Cole, S.T.; Leys, D.; Parish, T.; Munro, A.W. Characterization of active site structure in CYP121: a cytochrome P450 essential for viability of *Mycobacterium tuberculosis* H37Rv. *J Biol.*

Chem. **2008**, 283, 33406-33416.

- (15) Gondry M, Sauguet L, Belin P, Thai, R.; Amouroux, R.; Tellier, C.; Tiphile, K.; Jacquet, M.; Braud, S.; Courçon, M.; Masson, C.; Dubois, S.; Lautru, S.; Lecoq, A.; Hashimoto, S.; Genet, R.; Pernodet, J.L. Cyclodipeptide synthases are a family of tRNA- dependent peptide bond-forming enzymes. *Nat. Chem. Biol.* **2009**, 5, 414-420.
- (16) Vetting, M.W.; Hegde, S.S.; Blanchard, J.S. The structure and mechanism of the *Mycobacterium tuberculosis* cyclodityrosine synthetase. *Nat. Chem. Biol.* **2010**, 6, 797-799.
- (17) Belin, P.; Le Du, M.H.; Fielding A.; Lequin, O.; Jacquet, M.; Charbonnier, J.B.; Lecoq, A.; Thai, R; Courçon, M.; Masson, C.; Dugave, C.; Genet, R.; Pernodet, J.L.; Gony, M. Identification and structural basis of the reaction catalyzed by CYP121, an essential cytochrome P450 in *Mycobacterium tuberculosis*. *Proc. Natl. Acad. Sci. U.S.A.* **2009**, 106, 7426-7431.
- (18) Abd El-wahab, H.A.A.; Accietto, M.; Marino, L.B.; McLean, K.J.; Levy, C.W.; Abdel-Rahman, H.M.; El-Gendy, M.A.; Munro, A.W.; Aboraia, A.S.; Simons, C. Design, synthesis and evaluation against *Mycobacterium tuberculosis* of azole piperazine derivatives as dicyclotirosine (cYY) mimics. *Bioorg. Med. Chem.* **2018**, 26, 161-176.
- (19) Taban, I.M.; Elshihawy, H.E.A.E.; Torun, B.; Zucchini, B.; Williamson, C.J.; Altuwairigi, D.; Ngu, A.S.T.; McLean, K.J.; Levy, C.W.; Sood, S.; Marino, L.B.; Munro, A.W.; de Carvalho, L.P.S.; Simons, C. Novel aryl substituted pyrazoles as small molecule inhibitors of cytochrome P450 CYP121A1: synthesis and antimycobacterial evaluation. *J. Med. Chem.* **2017**, 60, 10257-10267.
- (20) Rondu, F.; Le Bihan, G.; Eang, X.; Lamouri, A.; Touboul, E.; Dive, G.; Bellahsene, T.; Pfeiffer, B.; Renard, P.; Guardiola-Lemaitre, B.; Manechez, D.; Penicaud, L.;

- Ktorza, A.; Godfroid, J.J. Design and synthesis of imidazoline derivatives active on glucose homeostasis in a rat model of type ii diabetes. 1. Synthesis and biological activities of N-benzyl-N'-(arylalkyl)-2-(4',5'-dihydro-1'H-imidazol-2'-yl)piperazines. *J. Med. Chem.* **1997**, *40*, 3793–3803.
- (21) Labute, P. LowModeMD - Implicit low-mode velocity filtering applied to conformational search of macrocycles and protein loops. *J. Chem. Inf. Model.* **2010**, *50*, 792–800.
- (22) *Molecular Operating Environment (MOE)*, 2015.10; Chemical Computing Group Inc., 1010 Sherbooke St. West, Suite #910, Montreal, QC, Canada, H3A 2R7, **2016**.
- (23) Stewart, J. J. P. Optimization of parameters for semiempirical methods. III Extension of PM3 to Be, Mg, Zn, Ga, Ge, As, Se, Cd, In, Sn, Sb, Te, Hg, Tl, Pb, and Bi. *J. Comput. Chem.* **1991**, *12*, 320–341.
- (24) Kallel, E. A.; Vangel, C.; Elbaum, D. Conformational analysis of 2-substituted piperazines. *Bioorg. Med. Chem. Lett.* **2016**, *26*, 3010–3013.
- (25) McLean, K.J.; Marshall, K.R.; Richmond, A.; Hunter, I.S.; Fowler, K.; Kieser, T.; Gurcha, S.S.; Besra, G.S.; Munro, A.W. Azole antifungals are potent inhibitors of cytochrome P450 mono-oxygenases and bacterial growth in mycobacteria and streptomycetes. *Microbiology* **2002**, *148*, 2937-2949.
- (26) Palomino, J.C.; Martin, A.; Camacho, M.; Guerra, H.; Swings, J.; Portaels, F. Resazurin microtiter assay plate: simple and inexpensive method for detection of drug resistance in *Mycobacterium tuberculosis*. *Antimicrob. Agents Chemother.* **2002**, *46*, 2720-2722.
- (27) Schrödinger Release 2019-1: Desmond Molecular Dynamics System, D. E. Shaw Research, New York, NY, 2019. Maestro-Desmond Interoperability Tools,

Schrödinger, New York, NY, 2019. [<https://www.schrodinger.com/Desmond/>]

(28) Bowers, K.J.; Chow, E.; Xu, H.; Dror, R.O.; Eastwood, M.P.; Gregersen, B.A.; Klepeis, J.L.; Kolossvary, I.; Moraes, M.A.; Sacerdoti, F.D.; Salmon, J.K.; Shan, Y.; Shaw, D.E. Scalable algorithms for molecular dynamics simulations on commodity clusters. Proceedings of the ACM/IEEE Conference on Supercomputing (SC06), Tampa, Florida, 2006, November 11-17.

(29) Husain, A.; Bhutani, R.; Kumar, D.; Shin, D. Synthesis and biological evaluation of novel substituted-imidazolidine derivatives. *J. Korean Chem. Soc.* **2013**, *57*, 227-233.

(30) Sharma, V.; Khan, M. S. Y. Synthesis of novel tetrahydroimidazole derivatives and studies for their biological properties. *Eur. J. Med. Chem.* **2001**, *36*, 651-658.

(31) Arish, D.; Nair, M. S. Synthesis, spectroscopic, antimicrobial, DNA binding and cleavage studies of some metal complexes involving symmetrical bidentate N, N donor Schiff base ligand. *Spectrochim. Acta A* **2011**, *82*, 191–199.

(32) Komatsu, H.; Ochiai, B.; Hino, T.; Endo, T. Thermally latent reaction of hemiacetal ester with epoxide controlled by Schiff-base-zinc chloride complexes with tunable catalytic activity. *J. Mol. Catal. A-Chem.* **2007**, *273*, 289–297.

(33) Al-Lami, A. K. Preparation and mesomorphic characterization of supramolecular hydrogen-bonded dimer liquid crystals. *Polycycl. Aromat. Comp.* **2016**, *36*, 197–212.

(34) Ouyang, X-M.; Fei, B-L.; Okamura, T-A.; Bu, H-W.; Sun, W-Y.; Tang, W-X.; Ueyama, N. Syntheses, crystal structures, and properties of four two-dimensional network complexes with multidentate bis(Schiff Base) ligands. *Eur. J. Chem.* **2003**, (4), 618-627.

(35) Buchs, B.; Godin, G.; Trachsel, A.; de Saint Laumer, J-Y.; Lehn, J-M.; Herrmann, A. Reversible aminal formation: controlling the evaporation of bioactive volatiles by dynamic combinatorial/covalent chemistry. *Eur. J. Org. Chem.* **2011**, (4) 681–695.